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(54) Title: GLYCOSIDASE ENZYMES			
(57) Abstract <p>A thermostable glycosidase enzymes derived from various <i>thermococcus</i>, <i>staphylothermus</i> and <i>pyrococcus</i> organisms is disclosed. The enzymes are produced from native or recombinant host cells and can be utilized in the food processing industry, pharmaceutical industry and in the textile industry, detergent industry and in the baking industry.</p>			

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GLYCOSIDASE ENZYMES

This application is a continuation-in-part of pending patent application 08/583,787 filed January 11, 1996.

This invention relates to newly identified polynucleotides, polypeptides encoded by such polynucleotides, the use of such polynucleotides and polypeptides, as well as the production and isolation of such polynucleotides and polypeptides. More particularly, the polynucleotides and polypeptides of the present invention has been putatively identified as glucosidases, α -galactosidases, β -galactosidases, β -mannosidases, β -mannanases, endoglucanases, and pullulanases.

The glycosidic bond of β -galactosides can be cleaved by different classes of enzymes: (i) phospho- β -galactosidases (EC3.2.1.85) are specific for a phosphorylated substrate generated via phosphoenolpyruvate phosphotransferase system (PTS)-dependent uptake; (ii) typical β -galactosidases (EC 3.2.1.23), represented by the *Escherichia coli* LacZ enzyme, which are relatively specific for β -galactosides; and (iii) β -glucosidases (EC 3.2.1.21) such as the enzymes of *Agrobacterium faecalis*, *Clostridium thermocellum*, *Pyrococcus furiosus* or *Sulfolobus solfataricus* (Day, A.G. and Withers, S.G., (1986) Purification and characterization of a β -glucosidase from *Alcaligenes faecalis*. *Can. J. Biochem. Cell. Biol.* 64, 914-922; Kengen, S.W.M., et al. (1993) *Eur. J. Biochem.*, 213, 305-312; Ait, N., Cruezet, N. and Cattaneo, J.

(1982) Properties of β -glucosidase purified from *Clostridium thermocellum*. *J. Gen. Microbiol.* 128, 569-577; Grogan, D.W. (1991) Evidence that β -galactosidase of *Sulfolobus solfataricus* is only one of several activities of a thermostable β -D-glycodiase. *Appl. Environ. Microbiol.* 57, 1644-1649). Members of the latter group, although highly specific with respect to the β -anomeric configuration of the glycosidic linkage, often display a rather relaxed substrate specificity and hydrolyse β -glucosides as well as β -fucosides and β -galactosides.

Generally, α -galactosidases are enzymes that catalyze the hydrolysis of galactose groups on a polysaccharide backbone or hydrolyze the cleavage of di- or oligosaccharides comprising galactose.

Generally, β -mannanases are enzymes that catalyze the hydrolysis of mannose groups internally on a polysaccharide backbone or hydrolyze the cleavage of di- or oligosaccharides comprising mannose groups. β -mannosidases hydrolyze non-reducing, terminal mannose residues on a mannose-containing polysaccharide and the cleavage of di- or oligosaccharides comprising mannose groups.

Guar gum is a branched galactomannan polysaccharide composed of β -1,4 linked mannose backbone with α -1,6 linked galactose sidechains. The enzymes required for the degradation of guar are β -mannanase, β -mannosidase and α -galactosidase. β -mannanase hydrolyses the mannose backbone internally and β -mannosidase hydrolyses non-reducing, terminal mannose residues. α -galactosidase hydrolyses α -linked galactose groups.

Galactomannan polysaccharides and the enzymes that degrade them have a variety of applications. Guar is commonly used as a thickening agent in food and is utilized in hydraulic fracturing in oil and gas recovery. Consequently, galactomannanases are industrially relevant for the degradation and modification of guar. Furthermore, a

need exists for thermostable galactomannases that are active in extreme conditions associated with drilling and well stimulation.

There are other applications for these enzymes in various industries, such as in the beet sugar industry. 20-30% of the domestic U.S. sucrose consumption is sucrose from sugar beets. Raw beet sugar can contain a small amount of raffinose when the sugar beets are stored before processing and rotting begins to set in. Raffinose inhibits the crystallization of sucrose and also constitutes a hidden quantity of sucrose. Thus, there is merit to eliminating raffinose from raw beet sugar. α -Galactosidase has also been used as a digestive aid to break down raffinose, stachyose, and verbascose in such foods as beans and other gassy foods.

β -Galactosidases which are active and stable at high temperatures appear to be superior enzymes for the production of lactose-free dietary milk products (Chaplin, M.F. and Bucke, C. (1990) In: *Enzyme Technology*, pp. 159-160, Cambridge University Press, Cambridge, UK). Also, several studies have demonstrated the applicability of β -galactosidases to the enzymatic synthesis of oligosaccharides via transglycosylation reactions (Nilsson, K.G.I. (1988) *Enzymatic synthesis of oligosaccharides*. *Trends Biotechnol.* 6, 156-264; Cote, G.L. and Tao, B.Y. (1990) *Oligosaccharide synthesis by enzymatic transglycosylation*. *Glycoconjugate J.* 7, 145-162). Despite the commercial potential, only a few β -galactosidases of thermophiles have been characterized so far. Two genes reported are β -galactoside-cleaving enzymes of the hyperthermophilic bacterium *Thermotoga maritima*, one of the most thermophilic organotrophic eubacteria described to date (Huber, R., Langworthy, T.A., König, H., Thomm, M., Woese, C.R., Sleytr, U.B. and Stetter, K.O. (1986) *T. maritima* sp. nov. represents a new genus of unique extremely thermophilic eubacteria growing up to 90°C, *Arch. Microbiol.* 144, 324-333) one of the most thermophilic organotrophic

eubacteria described to date. The gene products have been identified as a β -galactosidase and a β -glucosidase.

Pullulanase is well known as a debranching enzyme of pullulan and starch. The enzyme hydrolyzes α -1,6-glucosidic linkages on these polymers. Starch degradation for the production of sweeteners (glucose or maltose) is a very important industrial application of this enzyme. The degradation of starch is developed in two stages. The first stage involves the liquefaction of the substrate with α -amylase, and the second stage, or saccharification stage, is performed by β -amylase with pullulanase added as a debranching enzyme, to obtain better yields.

Endoglucanases can be used in a variety of industrial applications. For instance, the endoglucanases of the present invention can hydrolyze the internal β -1,4-glycosidic bonds in cellulose, which may be used for the conversion of plant biomass into fuels and chemicals. Endoglucanases also have applications in detergent formulations, the textile industry, in animal feed, in waste treatment, and in the fruit juice and brewing industry for the clarification and extraction of juices.

The polynucleotides and polypeptides of the present invention have been identified as glucosidases, α -galactosidases, β -galactosidases, β -mannosidases, β -mannanases, endoglucanases, and pullulanases as a result of their enzymatic activity.

In accordance with one aspect of the present invention, there are provided novel enzymes, as well as active fragments, analogs and derivatives thereof.

In accordance with another aspect of the present invention, there are provided isolated nucleic acid molecules encoding the enzymes of the present invention including mRNAs, cDNAs, genomic DNAs as well as active analogs and fragments of such enzymes.

In accordance with another aspect of the present invention there are provided isolated nucleic acid molecules encoding mature polypeptides expressed by the DNA contained in ATCC Deposit No. 97379.

In accordance with yet a further aspect of the present invention, there is provided a process for producing such polypeptides by recombinant techniques comprising culturing recombinant prokaryotic and/or eukaryotic host cells, containing a nucleic acid sequence of the present invention, under conditions promoting expression of said enzymes and subsequent recovery of said enzymes.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes, or polynucleotides encoding such enzymes for hydrolyzing lactose to galactose and glucose for use in the food processing industry, the pharmaceutical industry, for example, to treat intolerance to lactose, as a diagnostic reporter molecule, in corn wet milling, in the fruit juice industry, in baking, in the textile industry and in the detergent industry.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes for hydrolyzing guar gum (a galactomannan polysaccharide) to remove non-reducing terminal mannose residues. Further polysaccharides such as galactomannan and the enzymes according to the invention that degrade them have a variety of applications. Guar gum is commonly used as a thickening agent in food and also is utilized in hydraulic fracturing in oil and gas recovery. Consequently, mannanases are industrially relevant for the degradation and modification of guar gums. Furthermore, a need exists for thermostable mannanases that are active in extreme conditions associated with drilling and well stimulation.

In accordance with yet a further aspect of the present invention, there are also provided nucleic acid probes

comprising nucleic acid molecules of sufficient length to specifically hybridize to a nucleic acid sequence of the present invention.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes, or polynucleotides encoding such enzymes, for *in vitro* purposes related to scientific research, for example, to generate probes for identifying similar sequences which might encode similar enzymes from other organisms by using certain regions, i.e., conserved sequence regions, of the nucleotide sequence.

These and other aspects of the present invention should be apparent to those skilled in the art from the teachings herein.

Brief Description of the Drawings

The following drawings are illustrative of embodiments of the invention and are not meant to limit the scope of the invention as encompassed by the claims.

Figure 1 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of M11TL of the present invention. Sequencing was performed using a 378 automated DNA sequencer for all sequences of the present invention (Applied Biosystems, Inc.).

Figure 2 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of OC1/4V-33B/G.

Figure 3 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of F1-12G.

Figure 4 are illustrations of the full-length DNA and corresponding deduced amino acid sequence of 9N2-31B/G.

Figure 5 are illustrations of the full-length DNA and corresponding deduced amino acid sequence of MSB8-6G.

Figure 6 are illustrations of the full-length DNA and corresponding deduced amino acid sequence of AEDII12RA-18B/G.

Figure 7 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of GC74-22G.

Figure 8 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of VC1-7G1.

Figure 9 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of 37GP1.

Figure 10 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of 6GC2.

Figure 11 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of 6GP2.

Figure 12 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of 63GB1.

Figure 13 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of OC1/4V.

Figure 14 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of 6GP3.

Definitions

The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as intervening sequences (introns) between individual coding segments (exons).

A coding sequence is "operably linked to" another coding sequence when RNA polymerase will transcribe the two coding sequences into a single mRNA, which is then translated into a single polypeptide having amino acids derived from both coding sequences. The coding sequences need not be contiguous to one another so long as the expressed sequences ultimately process to produce the desired protein.

"Recombinant" enzymes refer to enzymes produced by recombinant DNA techniques; i.e., produced from cells transformed by an exogenous DNA construct encoding the desired enzyme. "Synthetic" enzymes are those prepared by chemical synthesis.

A DNA "coding sequence of" or a "nucleotide sequence encoding" a particular enzyme, is a DNA sequence which is

transcribed and translated into an enzyme when placed under the control of appropriate regulatory sequences.

Summary of the Invention

In accordance with an aspect of the present invention, there are provided isolated nucleic acids (polynucleotides) which encode for the mature enzymes having the deduced amino acid sequences of Figures 1-14 (SEQ ID NOS:15-28).

In accordance with another aspect of the present invention, there are provided isolated polynucleotides encoding the enzymes of the present invention. The deposited material is a mixture of genomic clones comprising DNA encoding an enzyme of the present invention. Each genomic clone comprising the respective DNA has been inserted into a pBluescript vector (Stratagene, La Jolla, CA). The deposit has been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852, USA, on December 13, 1995 and assigned ATCC Deposit No. 97379.

The deposit(s) have been made under the terms of the Budapest Treaty on the International Recognition of the deposit of micro-organisms for purposes of patent procedure. The strains will be irrevocably and without restriction or condition released to the public upon the issuance of a patent. These deposits are provided merely as convenience to those of skill in the art and are not an admission that a deposit be required under 35 U.S.C. §112. The sequences of the polynucleotides contained in the deposited materials, as well as the amino acid sequences of the polypeptides encoded thereby, are controlling in the event of any conflict with any description of sequences herein. A license may be required to make, use or sell the deposited materials, and no such license is hereby granted.

Detailed Description of the Invention

The polynucleotides of this invention were originally recovered from genomic gene libraries derived from the following organisms:

M11TL is a new species of *Desulfurococcus* isolated from Diamond Pool in Yellowstone National Park. The organism grows optimally at 85-88°C, pH 7.0 in a low salt medium containing yeast extract, peptone, and gelatin as substrates with a N₂/CO₂ gas phase.

OC1/4V is from the genus *Thermotoga*. The organism was isolated from Yellowstone National Park. It grows optimally at 75°C in a low salt medium with cellulose as a substrate and N₂ in gas phase.

Pyrococcus furiosus VC1 is from the genus *Pyrococcus*. VC1 was isolated from Vulcano, Italy. It grows optimally at 100°C in a high salt medium (marine) containing elemental sulfur, yeast extract, peptone and starch as substrates and N₂ in gas phase.

Staphylothermus marinus F1 is from the genus *Staphylothermus*. F1 was isolated from Vulcano, Italy. It grows optimally at 85°C, pH 6.5 in high salt medium (marine) containing elemental sulfur and yeast extract as substrates and N₂ in gas phase.

Thermococcus 9N-2 is from the genus *Thermococcus* 9N-2 was isolated from diffuse vent fluid in the East Pacific Rise. It is a strict anaerobe that grows optimally at 87°C.

Thermotoga maritima MSB8 is from the genus *Thermotoga*, and was isolated from Vulcano, Italy. MSB8 grows optimally at 85°C, pH 6.5 in a high salt medium (marine) containing starch and yeast extract as substrates and N₂ in gas phase.

Thermococcus alcaliphilus AEDIII12RA is from the genus *Thermococcus*. AEDIII12RA grows optimally at 85°C, pH 9.5 in a high salt medium (marine) containing polysulfides and yeast extract as substrates and N₂ in gas phase.

Thermococcus chitonophagus GC74 is from the genus *Thermococcus*. GC74 grows optimally at 85°C, pH 6.0 in a high salt medium (marine) containing chitin, meat extract, elemental sulfur and yeast extract as substrates and N₂ in gas

phase. AEPII 1a grows optimally at 85°C at pH 6.5 in marine medium under anaerobic conditions. It has many substrates.

[Add. descriptions of new organisms]

Accordingly, the polynucleotides and enzymes encoded thereby are identified by the organism from which they were isolated, and are sometimes hereinafter referred to as "M11TL" (Figure 1 and SEQ ID NOS:1 and 15), "OC1/4V-33B/G" (Figure 2 and SEQ ID NOS:2 and 16), "F1-12G" (Figure 3 and SEQ ID NOS:3 and 17), "9N2-31B/G" (Figure 4 and SEQ ID NOS:4 and 18), "MSB8" (Figure 5 and SEQ ID NOS:5 and 19), "AEDIII12RA-18B/G" (Figure 6 and SEQ ID NOS:6 and 20), "GC74-22G" (Figure 7 and SEQ ID NOS:7 and 21), "VC1-7G1" (Figure 8 and SEQ ID NOS:8 and 22), "37GP1" (Figure 9 and SEQ ID NOS:9 and 23), "6GC2" (Figure 10 and SEQ ID NOS:10 and 24), "6GP2" (Figure 11 and SEQ ID NOS:11 and 25), "AEPII 1a" (Figure 12 and SEQ ID NOS:12 and 26), "OC1/4V" (Figure 13 and SEQ ID NOS:13 and 27), and "6GP3" (Figure 14 and SEQ ID NOS:28).

The polynucleotides and polypeptides of the present invention show identity at the nucleotide and protein level to known genes and proteins encoded thereby as shown in Table 1.

Table 1

Clone	Gene/Protein with Closest Homology	Protein Identity	Nucleic Acid Identity
M11TL-29G	<i>Sulfolobus sulfataricus</i> DSM 1616/P1, β -galactosidase	51%	55%
OC1/4V-33B/G	<i>Caldocellum saccharolyticum</i> , β -glucosidase	52%	57%
<i>Staphylothermus marinus</i> F1-12G	<i>Bacillus polymyxa</i> , β -galactosidase	36%	48%

<i>Thermococcus</i> 9N2-31B/G	<i>Sulfolobus</i> <i>sulfataricus</i> ATCC 49255/MT4, β - galactosidase	51%	50%
<i>Thermotoga</i> <i>maritima</i> MSB8- 6G	<i>Clostridium</i> <i>thermocellum</i> bglB	45%	53%
<i>Thermococcus</i> AEDIII12RA-18B/G	<i>Bacillus polymyxa</i> , β -galactosidase	34%	48%
<i>Thermococcus</i> <i>chitonophagus</i> GC74-22G	<i>Sulfolobus</i> <i>sulfataricus</i> ATCC 49255/MT4, β - galactosidase	46%	54%
<i>Pyrococcus</i> <i>furious</i> VC1- 7G1	<i>Sulfolobus</i> <i>sulfataricus/MT-4</i> β -galactosidase	46.4%	52.5%
<i>Thermotoga</i> <i>maritima</i> α - galactosidase (6GC2)	<i>Pediococcus</i> <i>pentosaceus</i> α - galactosidase	49%	29%
<i>Thermotoga</i> <i>maritima</i> β - mannanase (6GP2)	<i>Aspergillus</i> <i>aculeatus</i> mannanase	56%	37%
AEP II 1a β - mannosidase (63GB1)	<i>Sulfolobus</i> <i>solfataricus</i> β - galactosidase	78%	56%
OC1/4V endoglucanase (33GP1)	<i>Clostridium</i> <i>thermocellum</i> endo- 1,4- β - endoglucanase	65%	43%
<i>Thermotoga</i> <i>maritima</i> pullulanase (6GP3)	<i>Caldocellum</i> <i>saccharolyticum</i> α - destrom 6 glucanohydralase	72	53
<i>Bankia gouldi</i> mix Endoglucanase (37GP1)	None available		

The polynucleotides and enzymes of the present invention show homology to each other as shown in Table 2.

Table 2

Clone	Gene/Protein with Closest Homology	Protein Identity	Nucleic Acid Identity
<i>Staphylothermus marinus</i> F1-12G	<i>Thermococcus</i> AEDIII12RA-18B/G, β -galactosidase, glucosidase	55%	57%
<i>Thermococcus</i> 9N2-31B/G	<i>Thermococcus chitonophagus</i> GC74-22G- glucosidase	74%	66%
<i>Pyrococcus furiosus</i> VC1-7G1	<i>Pyrococcus furiosus</i> VC1-7B/G β -galactosidase	46.4%	54%

All the clones identified in Tables 1 and 2 encode polypeptides which have α -glycosidase or β -glycosidase activity.

This invention, in addition to the isolated nucleic acid molecules encoding the enzymes of the present invention, also provide substantially similar sequences. Isolated nucleic acid sequences are substantially similar if: (i) they are capable of hybridizing under conditions hereinafter described, to the polynucleotides of SEQ ID NOS:1-8; (ii) or they encode DNA sequences which are degenerate to the polynucleotides of SEQ ID NOS:1-8. Degenerate DNA sequences encode the amino acid sequences of SEQ ID NOS:9-16, but have variations in the nucleotide coding sequences. As used herein, substantially similar refers to the sequences having similar identity to the sequences of the instant invention. The nucleotide sequences that are substantially the same can be identified by hybridization or by sequence comparison. Enzyme sequences that are substantially the same can be identified by one or more of the following: proteolytic digestion, gel electrophoresis and/or microsequencing.

One means for isolating the nucleic acid molecules encoding the enzymes of the present invention is to probe a gene library with a natural or artificially designed probe using art recognized procedures (see, for example: Current Protocols in Molecular Biology, Ausubel F.M. et al. (EDS.) Green Publishing Company Assoc. and John Wiley Interscience, New York, 1989, 1992). It is appreciated to one skilled in the art that the polynucleotides of SEQ ID NOS:1-14 or fragments thereof (comprising at least 12 contiguous nucleotides), are particularly useful probes. Other particular useful probes for this purpose are hybridizable fragments to the sequences of SEQ ID NOS:1-14 (i.e., comprising at least 12 contiguous nucleotides).

With respect to nucleic acid sequences which hybridize to specific nucleic acid sequences disclosed herein, hybridization may be carried out under conditions of reduced stringency, medium stringency or even stringent conditions. As an example of oligonucleotide hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at 45°C in a solution consisting of 0.9 M NaCl, 50 mM NaH₂PO₄, pH 7.0, 5.0 mM Na₂EDTA, 0.5% SDS, 10X Denhardt's, and 0.5 mg/mL polyriboadenylic acid. Approximately 2 X 10⁷ cpm (specific activity 4-9 X 10⁸ cpm/ug) of ³²P end-labeled oligonucleotide probe are then added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature in 1X SET (150 mM NaCl, 20 mM Tris hydrochloride, pH 7.8, 1 mM Na₂EDTA) containing 0.5% SDS, followed by a 30 minute wash in fresh 1X SET at T_m 10°C for the oligonucleotide probe. The membrane is then exposed to autoradiographic film for detection of hybridization signals.

Stringent conditions means hybridization will occur only if there is at least 90% identity, preferably at least 95% identity and most preferably at least 97% identity between the sequences. Further, it is understood that a section of

a 100 bps sequence that is 95 bps in length has 95% identity with the 1090 bps sequence from which it is obtained. See J. Sambrook et al., *Molecular Cloning, A Laboratory Manual*, 2d Ed., Cold Spring Harbor Laboratory (1989) which is hereby incorporated by reference in its entirety. Also, it is understood that a fragment of a 100 bps sequence that is 95 bps in length has 95% identity with the 100 bps sequence from which it is obtained.

As used herein, a first DNA (RNA) sequence is at least 70% and preferably at least 80% identical to another DNA (RNA) sequence if there is at least 70% and preferably at least a 80% or 90% identity, respectively, between the bases of the first sequence and the bases of the another sequence, when properly aligned with each other, for example when aligned by BLASTN.

"Identity" as the term is used herein, refers to a polynucleotide sequence which comprises a percentage of the same bases as a reference polynucleotide (SEQ ID NOS:1-8). For example, a polynucleotide which is at least 90% identical to a reference polynucleotide, has polynucleotide bases which are identical in 90% of the bases which make up the reference polynucleotide and may have different bases in 10% of the bases which comprise that polynucleotide sequence.

The present invention relates polynucleotides which differ from the reference polynucleotide such that the changes are silent changes, for example the change do not alter the amino acid sequence encoded by the polynucleotide. The present invention also relates to nucleotide changes which result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference polynucleotide. In a preferred aspect of the invention these polypeptides retain the same biological action as the polypeptide encoded by the reference polynucleotide.

It is also appreciated that such probes can be and are preferably labeled with an analytically detectable reagent to facilitate identification of the probe. Useful reagents include but are not limited to radioactivity, fluorescent dyes or enzymes capable of catalyzing the formation of a detectable product. The probes are thus useful to isolate complementary copies of DNA from other sources or to screen such sources for related sequences.

The polynucleotides of this invention were recovered from genomic gene libraries from the organisms listed in Table 1. For example, gene libraries can be generated in the Lambda ZAP II cloning vector (Stratagene Cloning Systems). Mass excisions can be performed on these libraries to generate libraries in the pBluescript phagemid. Libraries are thus generated and excisions performed according to the protocols/methods hereinafter described.

The excision libraries are introduced into the *E. coli* strain BW14893 F'kan1A. Expression clones are then identified using a high temperature filter assay. Expression clones encoding several glucanases and several other glycosidases are identified and repurified. The polynucleotides, and enzymes encoded thereby, of the present invention, yield the activities as described above.

The coding sequences for the enzymes of the present invention were identified by screening the genomic DNAs prepared for the clones having glucosidase or galactosidase activity.

An example of such an assay is a high temperature filter assay wherein expression clones were identified by use of high temperature filter assays using buffer Z (see recipe below) containing 1 mg/ml of the substrate 5-bromo-4-chloro-3-indolyl- β -D-glucopyranoside (XGLU) (Diagnostic Chemicals Limited or Sigma) after introducing an excision library into the *E. coli* strain BW14893 F'kan1A. Expression clones encoding XGLUases were identified and repurified from M11TL,

OC1/4V, *Pyrococcus furiosus* VC1, *Staphylothermus marinus* F1, *Thermococcus* 9N-2, *Thermotoga maritima* MSB8, *Thermococcus alcaliphilus* AEDII12RA, and *Thermococcus chitonophagus* GC74.

Z-buffer: (referenced in Miller, J.H. (1992) A Short Course in Bacterial Genetics, p. 445.)

per liter:

Na ₂ HPO ₄ ·7H ₂ O	16.1g
NaH ₂ PO ₄ ·7H ₂ O	5.5g
KCl	0.75g
MgSO ₄ ·7H ₂ O	0.246g
β-mercaptoethanol	2.7ml

Adjust pH to 7.0

High Temperature Filter Assay

- (1) The f factor f'kan (from *E. coli* strain CSH118) (1) was introduced into the pho-pnh-lac-strain BW14893 (2). BW13893 (2). The filamentous phage library was plated on the resulting strain, BW14893 F'kan. (Miller, J.H. (1992) A Short Course in Bacterial Genetics; Lee, K.S., Metcalf, et al., (1992) Evidence for two phosphonate degradative pathways in *Enterobacter Aerogenes*, *J. Bacteriol.*, 174:2501-2510.
- (2) After growth on 100 mm LB plates containing 100 µg/ml ampicillin, 80 µg/ml nethicillin and 1mM IPTG, colony lifts were performed using Millipore HATF membrane filters.
- (3) The colonies transferred to the filters were lysed with chloroform vapor in 150 mm glass petri dishes.
- (4) The filters were transferred to 100 mm glass petri dishes containing a piece of Whatman 3MM filter paper saturated with buffer.
 - (a) when testing for galactosidase activity (XGALase), 3MM paper was saturated with Z buffer containing 1 mg/ml XGAL (ChemBridge Corporation). After transferring filter bearing lysed colonies to

the glass petri dish, placed dish in oven at 80-85°C.

(b) when testing for glucosidase (XGLUase), 3MM paper was saturated with Z buffer containing 1 mg/ml XGLU. After transferring filter bearing lysed colonies to the glass petri dish, placed dish in oven at 80-85°C.

(5) 'Positives' were observed as blue spots on the filter membranes. Used the following filter rescue technique to retrieve plasmid from lysed positive colony. Used pasteur pipette (or glass capillary tube) to core blue spots on the filter membrane. Placed the small filter disk in an Eppendorf tube containing 20 µl water. Incubated the Eppendorf tube at 75°C for 5 minutes followed by vortexing to elute plasmid DNA off filter. This DNA was transformed into electrocompetent *E. coli* cells DH10B for *Thermatoga maritima* MSB8-6G, *Staphylothermus marinus* F1-12G, *Thermococcus AEDIII2RA-18B/G*, *Thermococcus chitonophagus* GC74-22G, M11T1 and OC1/4V. Electrocompetent BW14893 F'kan1A *E. coli* were used for *Thermococcus 9N2-31B/G*, and *Pyrococcus furiosus* VC1-7G1. Repeated filter-lift assay on transformation plates to identify 'positives'. Return transformation plates to 37°C incubator after filter lift to regenerate colonies. Inoculate 3 ml LB liquid containing 100 µg/ml ampicillin with repurified positives and incubate at 37°C overnight. Isolate plasmid DNA from these cultures and sequence plasmid insert. In some instances where the plates used for the initial colony lifts contained non-confluent colonies, a specific colony corresponding to a blue spot on the filter could be identified on a regenerated plate and repurified directly, instead of using the filter rescue technique.

Another example of such an assay is a variation of the high temperature filter assay wherein colony-laden filters

are heat-killed at different temperatures (for example, 105°C for 20 minutes) to monitor thermostability. The 3MM paper is saturated with different buffers (i.e., 100 mM NaCl, 5 mM MgCl₂, 100 mM Tris-Cl (pH 9.5)) to determine enzyme activity under different buffer conditions.

A β -glucosidase assay may also be employed, wherein Glcp β Np is used as an artificial substrate (aryl- β -glucosidase). The increase in absorbance at 405 nm as a result of p-nitrophenol (pNp) liberation was followed on a Hitachi U-1100 spectrophotometer, equipped with a thermostatted cuvette holder. The assays may be performed at 80°C or 90°C in closed 1-ml quartz cuvette. A standard reaction mixture contains 150 mM trisodium substrate, pH 5.0 (at 80°C), and 0.95 mM pNp derivative pNp = 0.561 mM⁻¹ • cm⁻¹). The reaction mixture is allowed to reach the desired temperature, after which the reaction is started by injecting an appropriate amount of enzyme (1.06 ml final volume).

1 U β -glucosidase activity is defined as that amount required to catalyze the formation of 1.0 μ mol pNp/min. D-cellobiose may also be used as a substrate.

An ONPG assay for β -galactosidase activity is described by Miller, J.H. (1992) A Short Course in Bacterial Genetics and Mill, J.H. (1992) Experiments in Molecular Genetics, the contents of which are hereby incorporated by reference in their entirety.

A quantitative fluorometric assay for β -galactosidase specific activity is described by : Youngman P., (1987) Plasmid Vectors for Recovering and Exploiting Tn917 Transpositions in *Bacillus* and other Gram-Positive Bacteria. In Plasmids: A Practical approach (ed. K. Hardy) pp 79-103. IRL Press, Oxford. A description of the procedure can be found in Miller (1992) p. 75-77, the contents of which are incorporated by reference herein in their entirety.

The polynucleotides of the present invention may be in the form of DNA which DNA includes cDNA, genomic DNA, and

synthetic DNA. The DNA may be double-stranded or single-stranded, and if single stranded may be the coding strand or non-coding (anti-sense) strand. The coding sequences which encodes the mature enzymes may be identical to the coding sequences shown in Figures 1-8 (SEQ ID NOS:1-8) or may be a different coding sequence which coding sequence, as a result of the redundancy or degeneracy of the genetic code, encodes the same mature enzymes as the DNA of Figures 1-14 (SEQ ID NOS:1-14).

The polynucleotide which encodes for the mature enzyme of Figures 1-14 (SEQ ID NOS:15-28) may include, but is not limited to: only the coding sequence for the mature enzyme; the coding sequence for the mature enzyme and additional coding sequence such as a leader sequence or a proprotein sequence; the coding sequence for the mature enzyme (and optionally additional coding sequence) and non-coding sequence, such as introns or non-coding sequence 5' and/or 3' of the coding sequence for the mature enzyme.

Thus, the term "polynucleotide encoding an enzyme (protein)" encompasses a polynucleotide which includes only coding sequence for the enzyme as well as a polynucleotide which includes additional coding and/or non-coding sequence.

The present invention further relates to variants of the hereinabove described polynucleotides which encode for fragments, analogs and derivatives of the enzymes having the deduced amino acid sequences of Figures 1-14 (SEQ ID NOS:15-28). The variant of the polynucleotide may be a naturally occurring allelic variant of the polynucleotide or a non-naturally occurring variant of the polynucleotide.

Thus, the present invention includes polynucleotides encoding the same mature enzymes as shown in Figures 1-14 (SEQ ID NOS:15-28) as well as variants of such polynucleotides which variants encode for a fragment, derivative or analog of the enzymes of Figures 1-14 (SEQ ID NOS:15-28). Such nucleotide variants include deletion

variants, substitution variants and addition or insertion variants.

As hereinabove indicated, the polynucleotides may have a coding sequence which is a naturally occurring allelic variant of the coding sequences shown in Figures 1-14 (SEQ ID NOS:1-14). As known in the art, an allelic variant is an alternate form of a polynucleotide sequence which may have a substitution, deletion or addition of one or more nucleotides, which does not substantially alter the function of the encoded enzyme.

Fragments of the full length gene of the present invention may be used as a hybridization probe for a cDNA or a genomic library to isolate the full length DNA and to isolate other DNAs which have a high sequence similarity to the gene or similar biological activity. Probes of this type preferably have at least 10, preferably at least 15, and even more preferably at least 30 bases and may contain, for example, at least 50 or more bases. The probe may also be used to identify a DNA clone corresponding to a full length transcript and a genomic clone or clones that contain the complete gene including regulatory and promotor regions, exons, and introns. An example of a screen comprises isolating the coding region of the gene by using the known DNA sequence to synthesize an oligonucleotide probe. Labeled oligonucleotides having a sequence complementary to that of the gene of the present invention are used to screen a library of genomic DNA to determine which members of the library the probe hybridizes to.

The present invention further relates to polynucleotides which hybridize to the hereinabove-described sequences if there is at least 70%, preferably at least 90%, and more preferably at least 95% identity between the sequences. The present invention particularly relates to polynucleotides which hybridize under stringent conditions to the hereinabove-described polynucleotides. As herein used,

the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences. The polynucleotides which hybridize to the hereinabove described polynucleotides in a preferred embodiment encode enzymes which either retain substantially the same biological function or activity as the mature enzyme encoded by the DNA of Figures 1-14 (SEQ ID NOS:1-14).

Alternatively, the polynucleotide may have at least 15 bases, preferably at least 30 bases, and more preferably at least 50 bases which hybridize to any part of a polynucleotide of the present invention and which has an identity thereto, as hereinabove described, and which may or may not retain activity. For example, such polynucleotides may be employed as probes for the polynucleotides of SEQ ID NOS:1-14, for example, for recovery of the polynucleotide or as a diagnostic probe or as a PCR primer.

Thus, the present invention is directed to polynucleotides having at least a 70% identity, preferably at least 90% identity and more preferably at least a 95% identity to a polynucleotide which encodes the enzymes of SEQ ID NOS:15-28 as well as fragments thereof, which fragments have at least 15 bases, preferably at least 30 bases and most preferably at least 50 bases, which fragments are at least 90% identical, preferably at least 95% identical and most preferably at least 97% identical under stringent conditions to any portion of a polynucleotide of the present invention.

The present invention further relates to enzymes which have the deduced amino acid sequences of Figures 1-14 (SEQ ID NOS:15-28) as well as fragments, analogs and derivatives of such enzyme.

The terms "fragment," "derivative" and "analog" when referring to the enzymes of Figures 1-14 (SEQ ID NOS:15-28) means enzymes which retain essentially the same biological function or activity as such enzymes. Thus, an analog

includes a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature enzyme.

The enzymes of the present invention may be a recombinant enzyme, a natural enzyme or a synthetic enzyme, preferably a recombinant enzyme.

The fragment, derivative or analog of the enzymes of Figures 1-14 (SEQ ID NOS:15-28) may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature enzyme is fused with another compound, such as a compound to increase the half-life of the enzyme (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature enzyme, such as a leader or secretory sequence or a sequence which is employed for purification of the mature enzyme or a proprotein sequence. Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

The enzymes and polynucleotides of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or enzyme present in a living animal is not isolated, but the same polynucleotide or enzyme, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or enzymes could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

The enzymes of the present invention include the enzymes of SEQ ID NOS:15-28 (in particular the mature enzyme) as well as enzymes which have at least 70% similarity (preferably at least 70% identity) to the enzymes of SEQ ID NOS:9-16 and more preferably at least 90% similarity (more preferably at least 90% identity) to the enzymes of SEQ ID NOS:15-28 and still more preferably at least 95% similarity (still more preferably at least 95% identity) to the enzymes of SEQ ID NOS:9-16 and also include portions of such enzymes with such portion of the enzyme generally containing at least 30 amino acids and more preferably at least 50 amino acids.

As known in the art "similarity" between two enzymes is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one enzyme to the sequence of a second enzyme.

A variant, i.e. a "fragment", "analog" or "derivative" polypeptide, and reference polypeptide may differ in amino acid sequence by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.

Among preferred variants are those that vary from a reference by conservative amino acid substitutions. Such substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr.

Most highly preferred are variants which retain the same biological function and activity as the reference polypeptide from which it varies.

Fragments or portions of the enzymes of the present invention may be employed for producing the corresponding full-length enzyme by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length enzymes. Fragments or portions of the polynucleotides of the present invention may be used to synthesize full-length polynucleotides of the present invention.

The present invention also relates to vectors which include polynucleotides of the present invention, host cells which are genetically engineered with vectors of the invention and the production of enzymes of the invention by recombinant techniques.

Host cells are genetically engineered (transduced or transformed or transfected) with the vectors of this invention which may be, for example, a cloning vector or an expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the genes of the present invention. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

The polynucleotides of the present invention may be employed for producing enzymes by recombinant techniques. Thus, for example, the polynucleotide may be included in any one of a variety of expression vectors for expressing an enzyme. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences, e.g., derivatives of SV40; bacterial plasmids; phage DNA; baculovirus; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and

pseudorabies. However, any other vector may be used as long as it is replicable and viable in the host.

The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease site(s) by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art.

The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned: LTR or SV40 promoter, the E. coli. lac or trp, the phage lambda P_L promoter and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. The expression vector also contains a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression.

In addition, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in E. coli.

The vector containing the appropriate DNA sequence as hereinabove described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein.

As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as E. coli, Streptomyces, Bacillus subtilis; fungal cells, such as yeast; insect cells such as Drosophila S2 and Spodoptera Sf9; animal cells such as CHO, COS or Bowes melanoma; adenoviruses; plant cells, etc. The selection of an appropriate host is deemed

to be within the scope of those skilled in the art from the teachings herein.

More particularly, the present invention also includes recombinant constructs comprising one or more of the sequences as broadly described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a sequence of the invention has been inserted, in a forward or reverse orientation. In a preferred aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and promoters are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example; Bacterial: pQE70, pQE60, pQE-9 (Qiagen), pD10, pSiX174, pBluescript II KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene); ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia); Eukaryotic: pSV2CAT, pOG44, pXT1, pSG (Stratagene) pSVK3, pBPV, pMSG, pSVL (Pharmacia). However, any other plasmid or vector may be used as long as they are replicable and viable in the host.

Promoter regions can be selected from any desired gene using CAT (chloramphenicol transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include lacI, lacZ, T3, T7, gpt, lambda P_R, P_L and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art.

In a further embodiment, the present invention relates to host cells containing the above-described constructs. The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a

bacterial cell. Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis, L., Dibner, M., Battey, J., *Basic Methods in Molecular Biology*, (1986)).

The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the enzymes of the invention can be synthetically produced by conventional peptide synthesizers.

Mature proteins can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters. Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook, et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor, N.Y., (1989), the disclosure of which is hereby incorporated by reference.

Transcription of the DNA encoding the enzymes of the present invention by higher eukaryotes is increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, a cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

Generally, recombinant expression vectors will include origins of replication and selectable markers permitting transformation of the host cell, e.g., the ampicillin resistance gene of E. coli and S. cerevisiae TRP1 gene, and a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence. Such

promoters can be derived from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), α -factor, acid phosphatase, or heat shock proteins, among others. The heterologous structural sequence is assembled in appropriate phase with translation initiation and termination sequences, and preferably, a leader sequence capable of directing secretion of translated enzyme. Optionally, the heterologous sequence can encode a fusion enzyme including an N-terminal identification peptide imparting desired characteristics, e.g., stabilization or simplified purification of expressed recombinant product.

Useful expression vectors for bacterial use are constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation initiation and termination signals in operable reading phase with a functional promoter. The vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desirable, provide amplification within the host. Suitable prokaryotic hosts for transformation include *E. coli*, *Bacillus subtilis*, *Salmonella typhimurium* and various species within the genera *Pseudomonas*, *Streptomyces*, and *Staphylococcus*, although others may also be employed as a matter of choice.

As a representative but nonlimiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, WI, USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period.

Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents, such methods are well known to those skilled in the art.

Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts, described by Gluzman, Cell, 23:175 (1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa and BHK cell lines. Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

The enzyme can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the mature

protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

The enzymes of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture). Depending upon the host employed in a recombinant production procedure, the enzymes of the present invention may be glycosylated or may be non-glycosylated. Enzymes of the invention may or may not also include an initial methionine amino acid residue.

β -galactosidase hydrolyzes lactose to galactose and glucose. Accordingly, the OC1/4V, 9N2-31B/G, AEDIII2RA-18B/G and F1-12G enzymes may be employed in the food processing industry for the production of low lactose content milk and for the production of galactose or glucose from lactose contained in whey obtained in a large amount as a by-product in the production of cheese. Generally, it is desired that enzymes used in food processing, such as the aforementioned β -galactosidases, be stable at elevated temperatures to help prevent microbial contamination.

These enzymes may also be employed in the pharmaceutical industry. The enzymes are used to treat intolerance to lactose. In this case, a thermostable enzyme is desired, as well. Thermostable β -galactosidases also have uses in diagnostic applications, where they are employed as reporter molecules.

Glucosidases act on soluble cellooligosaccharides from the non-reducing end to give glucose as the sole product. Glucanases (endo- and exo-) act in the depolymerization of cellulose, generating more non-reducing ends (endo-glucanases, for instance, act on internal linkages yielding cellobiose, glucose and cellooligosaccharides as products). β -glucosidases are used in applications where glucose is the

desired product. Accordingly, M11TL, F1-12G, GC74-22G and MSB8-6G (and OC1/4V, VC1-7G1, 9N2-31B/G and AEDII12RA18B/G) may be employed in a wide variety of industrial applications, including in corn wet milling for the separation of starch and gluten, in the fruit industry for clarification and equipment maintenance, in baking for viscosity reduction, in the textile industry for the processing of blue jeans, and in the detergent industry as an additive. For these and other applications, thermostable enzymes are desirable.

Antibodies generated against the enzymes corresponding to a sequence of the present invention can be obtained by direct injection of the enzymes into an animal or by administering the enzymes to an animal, preferably a nonhuman. The antibody so obtained will then bind the enzymes itself. In this manner, even a sequence encoding only a fragment of the enzymes can be used to generate antibodies binding the whole native enzymes. Such antibodies can then be used to isolate the enzyme from cells expressing that enzyme.

For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, 1975, *Nature*, 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., 1983, *Immunology Today* 4:72), and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole, et al., 1985, in *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96).

Techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce single chain antibodies to immunogenic enzyme products of this invention. Also, transgenic mice may be used to express humanized antibodies to immunogenic enzyme products of this invention.

Antibodies generated against the enzyme of the present invention may be used in screening for similar enzymes from other organisms and samples. Such screening techniques are known in the art, for example, one such screening assay is described in "Methods for Measuring Cellulase Activities", *Methods in enzymology*, Vol 160, pp. 87-116, which is hereby incorporated by reference in its entirety.

The present invention will be further described with reference to the following examples; however, it is to be understood that the present invention is not limited to such examples. All parts or amounts, unless otherwise specified, are by weight.

In order to facilitate understanding of the following examples certain frequently occurring methods and/or terms will be described.

"Plasmids" are designated by a lower case p preceded and/or followed by capital letters and/or numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

"Digestion" of DNA refers to catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinarily skilled artisan. For analytical purposes, typically 1 μ g of plasmid or DNA fragment is used with about 2 units of enzyme in about 20 μ l of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 μ g of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for

particular restriction enzymes are specified by the manufacturer. Incubation times of about 1 hour at 37°C are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion the reaction is electrophoresed directly on a polyacrylamide gel to isolate the desired fragment.

Size separation of the cleaved fragments is performed using 8 percent polyacrylamide gel described by Goeddel, D. et al., Nucleic Acids Res., 8:4057 (1980).

"Oligonucleotides" refers to either a single stranded polydeoxynucleotide or two complementary polydeoxynucleotide strands which may be chemically synthesized. Such synthetic oligonucleotides have no 5' phosphate and thus will not ligate to another oligonucleotide without adding a phosphate with an ATP in the presence of a kinase. A synthetic oligonucleotide will ligate to a fragment that has not been dephosphorylated.

"Ligation" refers to the process of forming phosphodiester bonds between two double stranded nucleic acid fragments (Maniatis, T., et al., Id., p. 146). Unless otherwise provided, ligation may be accomplished using known buffers and conditions with 10 units of T4 DNA ligase ("ligase") per 0.5 µg of approximately equimolar amounts of the DNA fragments to be ligated.

Unless otherwise stated, transformation was performed as described in the method of Graham, F. and Van der Eb, A., Virology, 52:456-457 (1973).

Example 1

Bacterial Expression and Purification of Glycosidase Enzymes

DNA encoding the enzymes of the present invention, SEQ ID NOS:1 through 8, were initially amplified from a pBluescript vector containing the DNA by the PCR technique using the primers noted herein. The amplified sequences were then inserted into the respective PQE vector listed beneath

the primer sequences, and the enzyme was expressed according to the protocols set forth herein. The 5' and 3' primer sequences for the respective genes are as follows:

Thermococcus AEDII12RA - 18B/G

5' CCGAGAATTCACTAAAGAGGGAGAAATTAACTATGGTGAATGCTATGATTGTC

(SEQ ID NO:29)

3' CGGAAGATCTTCATAGCTCCGAAGCCCATA (SEQ ID NO:30)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Bgl II.

OC1/4V-33B/G

5' CCGAGAATTCACTAAAGAGGGAGAAATTAACTATGATAAGAAGGTCCGATTTCC

(SEQ ID NO:31)

3' CGGAAGATCTTAAGATTAGAAATTCCCTT (SEQ ID NO:32)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Bgl II.

Thermococcus 9N2 - 31B/G

5' CCGAGAATTCACTAAAGAGGGAGAAATTAACTATGCTACCAGAAGGCTTCCTC

(SEQ ID NO:33)

3' CGGAGGTACCTCACCCAAAGTCCGAACCTTCTC (SEQ ID NO:34)

Vector: pQE30; and contains the following restriction enzyme sites 5' EcoRI and 3' KpnI.

Staphylothermus marinus F1 - 12G

5' CCGAGAATTCACTAAAGAGGGAGAAATTAACTATGATAAGGTTCTGATTAT

(SEQ ID NO:35)

3' CGGAAGATCTTATTGAGGTTCTTAATCC (SEQ ID NO:36)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Bgl II.

Thermococcus chitonophagus GC74 - 22G

5' CCGAGAATTCACTAAAGAGGGAGAAATTAACTATGCTTCCAGGAGAACTTTCTC

(SEQ ID NO:37)

3' CGGAGGATCCCTACCCCTCCTCTAAGATCTC (SEQ ID NO:38)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' BamHI.

M11TL

5' AATAATCTAGAGCATGCAATTCCCCAAAGACTTCATGATAG (SEQ ID NO:39)

3' AATAAAAGCTTACTGGATCAGTGTAAGATGCT (SEQ ID NO:40)

Vector: pQE70; and contains the following restriction enzyme sites 5' SphI and 3' Hind III.

Thermotoga maritima MSB8-6G

5' CCGACAATTGATTAAAGAGGAGAAATTAACATGGAAAGGATCGATGAAATT (SEQ ID NO:41)

3' CGGAGGTACCTCATGGTTGAATCTCTTCTC (SEQ ID NO:42)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' KpnI.

Pyrococcus furiosus VC1 - 7G1

5' CCGACAATTGATTAAAGAGGAGAAATTAACATGTTCCCTGAAAAGTTCCCTT (SEQ ID NO:43)

3' CGGAGGTACCTCATCCCCTCAGCAATTCTC (SEQ ID NO:44)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Kpn I.

Bankia gouldi endoglucanase (37GP1)

5' AATAAGGATCCGTTAGCGACGCTCGC

(SEQ ID NO:45)

3' AATAAAAGCTCCGGTTGTACAGCGTAATAGGC (SEQ ID NO:46)

Vector: pQE52; and contains the following restriction enzyme sites 5' Bam HI and 3' Hind III.

Thermotoga maritima α -galactosidase (6GC2)

5' TTTATTGAATTCTTAAAGAGGAGAAATTAACATGATCTGTGTGGAAATATTGGAAAG

(SEQ ID NO:47)

3' TCTATAAAGCTTCTTCAATTCTCTCACCCCTTCTCGTAGAAG (SEQ ID NO:48)

Vector: pQET; and contains the following restriction enzyme sites 5' EcoRI and 3' Hind III.

Thermotoga maritima β -mannanase (6GP2)

5' TTTATTCAATTGATTAAAGAGGGAGAAATTAACTATGGGGATTGGTGGCGACGAC

(SEQ ID NO:49)

3' TTTATTAAGCTTATCTTTCATATTACACATACCTCC (SEQ ID NO:50)

Vector: pQEt; and contains the following restriction enzyme sites 5' Hind III and 3' EcoRI.

AEP II 1a β -mannanase (63GB1)

5' TTTATTGAATTCAATTAAAGAGGGAGAAATTAACTATGCTACCAGAAGAGTTCTATGGGC

(SEQ ID NO:51)

3' TTTATTAAGCTTCTCATCAACGGCTATGGTCTTCATTTC (SEQ ID NO:52)

Vector: pQE_t; and contains the following restriction enzyme sites 5' Hind III and 3' EcoRI.

OC1/4V endoglucanase (33GP1)

5' AAAAAACAATTGAATTCAATTAAAGAGGGAGAAATTAACTATGGTAGAAAGACACTTCAGATATGTTCTT

(SEQ ID NO:53)

3' TTTTCGGATCCAATTCTTCATTACTCTTGCCTG (SEQ ID NO:54)

Vector: pQE_t; and contains the following restriction enzyme sites 5' BamHI and 3' EcoRI.

Thermotoga maritima pullulanase (6GP3)

5' TTTTGGAAATTCAATTAAAGAGGGAGAAATTAACTATGGAACGTGATCATAGAAGGTTAC

(SEQ ID NO:55)

3' ATAAGAACGCTTCACTCTGTACAGAACGTACGC (SEQ ID NO:56)

Vector: pQE_t; and contains the following restriction enzyme sites 5' EcoRI and 3' Hind III.

The restriction enzyme sites indicated correspond to the restriction enzyme sites on the bacterial expression vector indicated for the respective gene (Qiagen, Inc. Chatsworth, CA). The pQE vector encodes antibiotic resistance (Amp'), a bacterial origin of replication (ori), an IPTG-regulatable

promoter operator (P/O), a ribosome binding site (RBS), a 6-His tag and restriction enzyme sites.

The pQE vector was digested with the restriction enzymes indicated. The amplified sequences were ligated into the respective pQE vector and inserted in frame with the sequence encoding for the RBS. The ligation mixture was then used to transform the *E. coli* strain M15/pREP4 (Qiagen, Inc.) by electroporation. M15/pREP4 contains multiple copies of the plasmid pREP4, which expresses the lacI repressor and also confers kanamycin resistance (Kan'). Transformants were identified by their ability to grow on LB plates and ampicillin/kanamycin resistant colonies were selected. Plasmid DNA was isolated and confirmed by restriction analysis. Clones containing the desired constructs were grown overnight (O/N) in liquid culture in LB media supplemented with both Amp (100 ug/ml) and Kan (25 ug/ml). The O/N culture was used to inoculate a large culture at a ratio of 1:100 to 1:250. The cells were grown to an optical density 600 (O.D.⁶⁰⁰) of between 0.4 and 0.6. IPTG ("Isopropyl-B-D-thiogalacto pyranoside") was then added to a final concentration of 1 mM. IPTG induces by inactivating the lacI repressor, clearing the P/O leading to increased gene expression. Cells were grown an extra 3 to 4 hours. Cells were then harvested by centrifugation.

The primer sequences set out above may also be employed to isolate the target gene from the deposited material by hybridization techniques described above.

Example 2

Isolation of A Selected Clone From the Deposited genomic clones

A clone is isolated directly by screening the deposited material using the oligonucleotide primers set forth in Example 1 for the particular gene desired to be isolated. The specific oligonucleotides are synthesized

using an Applied Biosystems DNA synthesizer. The oligonucleotides are labeled with ^{32}P -ATP using T4 polynucleotide kinase and purified according to a standard protocol (Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, Cold Spring, NY, 1982). The deposited clones in the pBluescript vectors may be employed to transform bacterial hosts which are then plated on 1.5% agar plates to the density of 20,000-50,000 pfu/150 mm plate. These plates are screened using Nylon membranes according to the standard screening protocol (Stratagene, 1993). Specifically, the Nylon membrane with denatured and fixed DNA is prehybridized in 6 x SSC, 20 mM NaH₂PO₄, 0.4% SDS, 5 x Denhardt's 500 $\mu\text{g}/\text{ml}$ denatured, sonicated salmon sperm DNA; and 6 x SSC, 0.1% SDS. After one hour of prehybridization, the membrane is hybridized with hybridization buffer 6xSSC, 20 mM NaH₂PO₄, 0.4% SDS, 500 $\mu\text{g}/\text{ml}$ denatured, sonicated salmon sperm DNA with 1×10^6 cpm/ml ^{32}P -probe overnight at 42°C. The membrane is washed at 45-50°C with washing buffer 6 x SSC, 0.1% SDS for 20-30 minutes dried and exposed to Kodak X-ray film overnight. Positive clones are isolated and purified by secondary and tertiary screening. The purified clone is sequenced to verify its identity to the primer sequence.

Once the clone is isolated, the two oligonucleotide primers corresponding to the gene of interest are used to amplify the gene from the deposited material. A polymerase chain reaction is carried out in 25 μl of reaction mixture with 0.5 μg of the DNA of the gene of interest. The reaction mixture is 1.5-5 mM MgCl₂, 0.01% (w/v) gelatin, 20 μM each of dATP, dCTP, dGTP, dTTP, 25 pmol of each primer and 0.25 Unit of Taq polymerase. Thirty five cycles of PCR (denaturation at 94°C for 1 min; annealing at 55°C for 1 min; elongation at 72°C for 1 min) are performed with the Perkin-Elmer Cetus automated thermal cycler. The amplified product is analyzed by agarose gel electrophoresis and the

DNA band with expected molecular weight is excised and purified. The PCR product is verified to be the gene of interest by subcloning and sequencing the DNA product. The ends of the newly purified genes are nucleotide sequenced to identify full length sequences. Complete sequencing of full length genes is then performed by Exonuclease III digestion or primer walking.

Example 3

Screening for Galactosidase Activity

Screening procedures for α -galactosidase protein activity may be assayed for as follows:

Substrate plates were provided by a standard plating procedure. Dilute XL1-Blue MRF E coli host of (Stratagene Cloning Systems, La Jolla, CA) to O.D.₆₀₀ = 1.0 with NYZ media. In 15 ml tubes, inoculate 200 μ l diluted host cells with phage. Mix gently and incubate tubes at 37 °C for 15 min. Add approximately 3.5 ml LB top agarose (0.7%) containing 1mM IPTG to each tube and pour onto all NYZ plate surface. Allow to cool and incubate at 37 °C overnight. The assay plates are obtained as substrate p-Nitrophenyl α -galactosidase (Sigma) (200 mg/100 ml) (100 mM NaCl, 100 mM Potassium-Phosphate) 1% (w/v) agarose. The plaques are overlayed with nitrocellulose and incubated at 4 °C for 30 minutes whereupon the nitrocellulose is removed and overlayed onto the substrate plates. The substrate plates are then incubated at 70 °C for 20 minutes.

Example 4

Screening of Clones for Mannanase Activity

A solid phase screening assay was utilized as a primary screening method to test clones for β -mannanase activity.

A culture solution of the Y1090-E. coli host strain (Stratagene Cloning Systems, La Jolla, CA) was diluted to

O.D.₆₀₀=1.0 with NZY media. The amplified library from *Thermotoga maritima* lambda gt11 library was diluted in SM (phage dilution buffer): 5 x 10⁷ pfu/μl diluted 1:1000 then 1:100 to 5 x 10² pfu/μl. Then 8 μl of phage dilution (5 x 10² pfu/μl) was plated in 200 μl host cells. They were then incubated in 15 ml tubes at 37 °C for 15 minutes.

Approximately 4 ml of molten, LB top agarose (0.7%) at approximately 52 °C was added to each tube and the mixture was poured onto the surface of LB agar plates. The agar plates were then incubated at 37 °C for five hours. The plates were replicated and induced with 10 mM IPTG-soaked Duralon-UV™ nylon membranes (Stratagene Cloning Systems, La Jolla, CA) overnight. The nylon membranes and plates were marked with a needle to keep their orientation and the nylon membranes were then removed and stored at 4 °C.

An Azo-galactomannan overlay was applied to the LB plates containing the lambda plaques. The overlay contains 1% agarose, 50 mM potassium-phosphate buffer pH 7, 0.4% Azocarob-galactomannan. (Megazyme, Australia). The plates were incubated at 72 °C. The Azocarob-galactomannan treated plates were observed after 4 hours then returned to incubation overnight. Putative positives were identified by clearing zones on the Azocarob-galactomannan plates. Two positive clones were observed.

The nylon membranes referred to above, which correspond to the positive clones were retrieved, oriented over the plate and the portions matching the locations of the clearing zones for positive clones were cut out. Phage was eluted from the membrane cut-out portions by soaking the individual portions in 500 μl SM (phage dilution buffer) and 25 μl CHCl₃.

Example 5

Screening of Clones for Mannosidase Activity

A solid phase screening assay was utilized as a primary screening method to test clones for β -mannosidase activity.

A culture solution of the Y1090-E. coli host strain (Stratagene Cloning Systems, La Jolla, CA) was diluted to $O.D_{600}=1.0$ with NZY media. The amplified library from AEPII 1a lambda gt11 library was diluted in SM (phage dilution buffer): 5×10^7 pfu/ μ l diluted 1:1000 then 1:100 to 5×10^2 pfu/ μ l. Then 8 μ l of phage dilution (5×10^2 pfu/ μ l) was plated in 200 μ l host cells. They were then incubated in 15 ml tubes at 37 °C for 15 minutes.

Approximately 4 ml of molten, LB top agarose (0.7%) at approximately 52 °C was added to each tube and the mixture was poured onto the surface of LB agar plates. The agar plates were then incubated at 37 °C for five hours. The plates were replicated and induced with 10 mM IPTG-soaked Duralon-Uv™ nylon membranes (Stratagene Cloning Systems, La Jolla, CA) overnight. The nylon membranes and plates were marked with a needle to keep their orientation and the nylon membranes were then removed and stored at 4 °C.

A p-nitrophenyl- β -D-manno-pyranoside overlay was applied to the LB plates containing the lambda plaques. The overlay contains 1% agarose, 50 mM potassium-phosphate buffer pH 7, 0.4% p-nitrophenyl- β -D-manno-pyranoside. (Megazyme, Australia). The plates were incubated at 72 °C. The p-nitrophenyl- β -D-manno-pyranoside treated plates were observed after 4 hours then returned to incubation overnight. Putative positives were identified by clearing zones on the p-nitrophenyl- β -D-manno-pyranoside plates. Two positive clones were observed.

The nylon membranes referred to above, which correspond to the positive clones were retrieved, oriented over the plate and the portions matching the locations of the clearing zones for positive clones were cut out. Phage was eluted from the membrane cut-out portions by soaking

the individual portions in 500 μ l SM (phage dilution buffer) and 25 μ l CHCl₃.

Example 6

Screening for Pullulanase Activity

Screening procedures for pullulanase protein activity may be assayed for as follows:

Substrate plates were provided by a standard plating procedure. Host cells are diluted to O.D.₆₀₀ = 1.0 with NZY or appropriate media. In 15 ml tubes, inoculate 200 μ l diluted host cells with phage. Mix gently and incubate tubes at 37 °C for 15 min. Add approximately 3.5 ml LB top agarose (0.7%) is added to each tube and the mixture is plated, allowed to cool, and incubated at 37°C for about 28 hours. Overlays of 4.5 mls of the following substrate are poured:

100 ml total volume

0.5g	Red Pullulan Red (Megazyme, Australia)
1.0g	Agarose
5ml	Buffer (Tris-HCL pH 7.2 @ 75 °C)
2ml	5M NaCl
5ml	CaCl ₂ (100mM)
85ml	dH ₂ O

Plates are cooled at room temperature, and thenm incubated at 75°C for 2 hours. Positives are observed as showing substrate degradation.

Example 7

Screening for Endoglucanase Activity

Screening procedures for endoglucanase protein activity may be assayed for as follows:

1. The gene library is plated onto 6 LB/GelRite/0.1% CMC/NZY agar plates (~4,800 plaque forming units/plate) in E.coli host with LB agarose as top agarose. The plates are incubated at 37°C overnight.

2. Plates are chilled at 4°C for one hour.
3. The plates are overlayed with Duralon membranes (Stratagene) at room temperature for one hour and the membranes are oriented and lifted off the plates and stored at 4°C.
4. The top agarose layer is removed and plates are incubated at 37°C for ~3 hours.
5. The plate surface is rinsed with NaCl.
6. The plate is stained with 0.1% Congo Red for 15 minutes.
7. The plate is destained with 1M NaCl.
8. The putative positives identified on plate are isolated from the Duralon membrane (positives are identified by clearing zones around clones). The phage is eluted from the membrane by incubating in 500µl SM + 25µl CHCl, to elute.
9. Insert DNA is subcloned into any appropriate cloning vector and subclones are reassayed for CMCase activity using the following protocol:
 - i) Spin 1ml overnight miniprep of clone at maximum speed for 3 minutes.
 - ii) Decant the supernatant and use it to fill "wells" that have been made in an LB/GelRite/0.1% CMC plate.
 - iii) Incubate at 37°C for 2 hours.
 - iv) Stain with 0.1% Congo Red for 15 minutes.
 - v) Destain with 1M NaCl for 15 minutes.
 - vi) Identify positives by clearing zone around clone.

Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, within the scope of the appended claims, the invention may be practiced otherwise than as particularly described.

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising a member selected from the group consisting of:
 - (a) a polynucleotide having at least a 70% identity to a polynucleotide encoding an enzyme comprising amino acid sequences set forth in SEQ ID NOS:15-28;
 - (b) a polynucleotide which is complementary to the polynucleotide of (a); and
 - (c) a polynucleotide comprising at least 15 bases of the polynucleotide of (a) or (b).
2. The polynucleotide of Claim 1 wherein the polynucleotide is DNA.
3. The polynucleotide of Claim 1 wherein the polynucleotide is RNA.
4. The polynucleotide of Claim 2 which encodes an enzyme comprising an amino acid sequence which a member selected from the group
 - (a) according to SEQ ID NO:15;
 - (b) according to SEQ ID NO:16;
 - (c) according to SEQ ID NO:17;
 - (d) according to SEQ ID NO:18;
 - (e) according to SEQ ID NO:19;
 - (f) according to SEQ ID NO:20;
 - (g) according to SEQ ID NO:21;
 - (h) according to SEQ ID NO:22;
 - (i) according to SEQ ID NO:23;
 - (j) according to SEQ ID NO:24;
 - (k) according to SEQ ID NO:25;
 - (l) according to SEQ ID NO:26;
 - (m) according to SEQ ID NO:27; and
 - (n) according to SEQ ID NO:28.

5. An isolated polynucleotide comprising a member selected from the group consisting of:

(a) a polynucleotide having at least a 70% identity to a polynucleotide encoding an enzyme encoded by the DNA contained in ATCC Deposit No. 97379, wherein said enzyme is selected from the group consisting of M11TL, OC1/4V, F1-12G, 9N2-31B/G, MSB8-6G, AEDIII12RA-18B/G, GC74-22G and VC1-7G1;

(b) a polynucleotide complementary to the polynucleotide of (a); and

(c) a polynucleotide comprising at least 15 bases of the polynucleotide of (a) and (b).

6. A vector comprising the DNA of Claim 2.

7. A host cell comprising the vector of Claim 6.

8. A process for producing a polypeptide comprising: expressing from the host cell of Claim 7 a polypeptide encoded by said DNA.

9. A process for producing a cell comprising: transforming or transfecting the cell with the vector of Claim 6 such that the cell expresses the polypeptide encoded by the DNA contained in the vector.

10. An enzyme comprising a member selected from the group consisting of:

(a) an enzyme comprising an amino acid sequence which is at least 70% identical to the amino acid sequence set forth in SEQ ID NOS:15-28; and

(b) an enzyme which comprises at least 30 amino acid residues to the enzyme of (a).

11. A method for generating glucose from soluble celooligosaccharides comprising:
administering an effective amount of an enyzme selected from the group consisting of an enzyme having the amino acid sequence set forth in SEQ ID NOS:15-28.

M11TL GLYCOSIDASE - 29G
COMPLETE GENE SEQUENCE - 9/95

1	TTG AAA TTC CGG AAA GAC TTC ATT ATA TAC TAA TGA TTT TCA CGG TTT GAA GCG	60
2	Met Lys Tyr Pro Lys Asp Phe Met Ile Lys Tyr Asp Ser Pro Pro Glu Pro Glu Ala	60
60	CTG ATT CGG CGG TCC GAG GAT CGG AAT ACT GAT TGC TGG GCA TGG CGG CAT GAT CGG GAG	120
61	Gly Ile Pro Gly Ser Glu Asp Pro Asp Ser Asp Thr Trp Val Thr Val Val His Asp Pro Glu	120
121	AAC ACA GCA GCT GGA CTA CGC AGC CGC GAT TTT CGG GAG AAC GGC CCA GCT TAC TGC AAT	180
122	Asn Thr Ala Ala Gly Leu Val Ser Gly Asp Phe Pro Glu Asn Glu Pro Gly Tyr Thr Asn	180
181	TTA AAC CAA AAT GAC CAC GAC CGC CCT GAG AAG CTG CGG CTT AAC ACT ATT AGA GCA CGG	240
182	Leu Asn Gln Asn Asp His Asp Leu Ala Glu Leu Gly Val Asn Thr Ile Arg Val Gly	240
241	CTT GAG TCG ACT AGG ATT TTT CCA AAG CCA ACT TTC AAT GTT AAA GTC CCT GTA GAG AGA	300
242	Val Glu Trp Ser Arg Ile Phe Pro Lys Pro Thr Phe Asn Val Lys Val Val Pro Val Glu Arg	300
301	GAT GAG AAC CGC AGC ATT CTT CAC GTA GAT GTC GAT GAT AAA GCG GTT GAA AGA CTT GAT	360
302	Asp Glu Asn Gly Ser Ile Val His Val Asp Val Asp Lys Ala Val Glu Arg Leu Asp	360
361	GAA TTA CGC AAC AAG GAG CGC CTA AAC CAT TAC GTA GAA ATG TAT AAA GAC TCG GTT GAA	420
362	Glu Leu Ala Asn Lys Glu Ala Val Asn His Tyr Val Glu Met Tyr Lys Asp Trp Val Glu	420
421	AGA CGT AGA AAA CTT ATA CTC AAT TTA TAC CAT TGG CCC CTG CCT CTC TGG CTT CAC AAC	480
422	Arg Gly Arg Lys Leu Ile Leu Asn Leu Tyr His Trp Pro Leu Pro Trp Leu His Asn	480
481	CCA ATC ATG GTG AGA AGA ATG CGC CCG GAC AGA CGC CCC TCA CGC TGG CTT AAC GAG GAG	540
482	Pro Ile Met Val Arg Arg Met Gly Pro Asp Arg Ala Pro Ser Gly Trp Leu Asn Glu Glu	540
541	TCC GTG GTG GAG TTT CGC AAA TAC CGC GCA TAC ATT GCT TGG AAA ATG CGC GAG CTA CCT	600
542	Ser Val Val Glu Phe Ala Lys Tyr Ile Ala Trp Lys Met Gly Glu Leu Pro	600
601	GTT ATG TGG AGC ACC ATG AAC GAA CCC AAC GTC GTT TAT GAG CAA GGA TAC ATG TTC GTT	660
602	Val Met Trp Ser Thr Met Asn Glu Pro Asn Val Val Tyr Glu Gln Gly Tyr Met Phe Val	660
661	AAA CGG CGT TTC CCA CCC CGC TAC TTG AGT TTG GAA GCT GCT GAT AAG GCC AGG AGA AAT	720
662	Lys Gly Gly Phe Pro Pro Gly Tyr Leu Ser Leu Glu Ala Ala Asp Lys Ala Arg Arg Asn	720
721	ATG ATC CAG GCT CAT GCA CGG CGC TAT GAC AAT ATT AAA CGC TTC AGT AAG AAA CCT GTT	780
722	Met Ile Gln Ala His Ala Arg Ala Tyr Asp Asn Ile Lys Arg Phe Ser Lys Lys Pro Val	780
781	GGA CTA ATA TAC GCT TTC CAA TGG TTC GAA CTA TTA GAG GGT CCA GCA GAA GTA TTT GAT	840
782	Gly Leu Ile Tyr Ala Phe Gln Trp Phe Glu Leu Leu Glu Gly Pro Ala Glu Val Phe Asp	840
841	AAC TTT AAG AGC TCT AAG TTA TAC TAT TTC ACA GAC ATA GTA TCG AAG GGT AGT TCA ATC	900
842	Lys Phe Lys Ser Ser Lys Leu Tyr Phe Thr Asp Ile Val Ser Lys Gly Ser Ser Ile	900
901	ATC AAT GTT GAA TAC AGG AGA GAT CTT CGC AAT AGG CTA GAC TGG TTG CGC GTT AAC TAC	960
902	Ile Asn Val Glu Tyr Arg Arg Asp Leu Ala Asn Arg Leu Asp Trp Leu Gly Val Asn Tyr	960
961	TAT AGC CGT TTA GTC TAC AAA ATC GTC GAT GAC AAA CCT ATA ATC CTG CAC CGC TAT CGA	1020
962	Tyr Ser Arg Leu Val Tyr Ile Val Asp Asp Lys Pro Ile Ile Leu His Gly Tyr Gly	1020
1021	TTC CTT TGT ACA CCT CGG CGG ATC AGC CGG GCT GAA AAT CCT TGT ACC GAT TTT CGG TGG	1080
1022	Phe Leu Cys Thr Pro Gly Ile Ser Pro Ala Glu Asn Pro Cys Ser Asp Phe Gly Trp	1080
1081	GAG GTG TAT CCT GAA CGG CTC TAC CTA CTT CTA AAA GAA CTT TAC AAC CGA TAC CGG CTA	1140
1082	Glu Val Tyr Pro Glu Gly Leu Tyr Leu Leu Lys Glu Leu Tyr Asn Arg Tyr Gly Val	1140
1141	GAC TTC ATC CGC ACC GAC AAC GGT CTT TCA GAC AGC AAG GAT CGG TTG AGA CGC GCA TAC	1200
1142	Asp Leu Ile Val Thr Glu Asn Gly Val Ser Asp Ser Arg Asp Ala Leu Arg Pro Ala Tyr	1200
1201	CTG GTC TGG CAT CTT TAC AGC GTC TGG AAA CGC CGT AAC GAC CGC ATT CGC CTC AAA CGC	1260
1202	Leu Val Ser His Val Tyr Ser Val Trp Lys Ala Ala Asn Glu Glu Ile Pro Val Lys Gly	1260
1261	TAC CTC GAC TGT AGC TTT GCA GAT AAT TAC GAG TGG CGT GAG CGC TTT AGC AAC AAA TTT	1320
1262	Tyr Leu His Trp Ser Leu Thr Asp Asn Tyr Glu Trp Ala Glu Gly Phe Arg Glu Lys Phe	1320

Figure 1

1441	CCT TTA GGT ATG GTC GAA TTC AAA AGT AAA AAA AGG TAT CTC GAA GAA A AGC GCG TTA ATG	1442
441	Pro Leu Val Ser Val Ala Thr Tyr Tyr Lys Arg Tyr Leu Asp Ile Val	442
1443	TTC CGG GAG ATG GCA ACC GAT AAA GUA AAA CGG GAT GAG CTC GAG GAT CTC ACA CGG ATG	1444
443	Phe Arg Glu Leu Ala Thr Ile Arg Gln Ile Pro Asp Glu Ile Gln Ile Leu Thr Lys Phe	444
1445	CAG TAA 1446	
445	GTC End 482	

Figure 1 (Continued)

OC1/4 GLYCOSIDASE - 33G/B
COMPLETE GENE SEQUENCE - 9/95

1	ATG ATA AGA AGG TCC GAT TTT CCA AAA GAT TTT ATC TTC GGA AGG GGT AGC GCA GCA TAC	60
1	Met Ile Arg Arg Ser Asp Phe Phe Lys Asp Phe Ile Phe Gly Thr Ala Thr Ala Ala Tyr	20
61	CTG ATT GAA GGT GCA GCA AAC GAA GAT GGC AGA GGG CCA TCA ATT TCG GAT GTC TTT TCA	120
61	Gln Ile Glu Gly Ala Ala Asn Glu Asp Gly Arg Gly Pro Ser Ile Thr Asp Val Phe Ser	40
121	CAC ACG CCT GGC AAA ACC CTG AAC GGT GAC ACA GGA GAC GTT GCG TCT GAC CAT TAT CAC	180
41	Ile Thr Pro Gly Lys Thr Leu Asn Gly Asp Thr Gly Asp Val Ala Cys Asp His Tyr His	60
181	CGA TAC AAG GAA GAT ATC CAG CTG ATG AAA GAA ATA GGG TTA GAC CCT TAC AGG TTC TCT	240
61	Arg Tyr Lys Glu Asp Ile Gln Leu Met Lys Glu Ile Gly Leu Asp Ala Tyr Arg Phe Ser	80
241	ATC TCC TGG CCC AGA ATT ATG CCA GAT GGG AAG AAC ATC AAC CAA AAC CCT CTG GAT TTC	300
81	Ile Ser Trp Pro Arg Ile Met Pro Asp Gly Lys Asn Ile Asn Gln Lys Gly Val Asp Phe	100
301	TAC AAC AGA CTC GTT GAT GAG CTT TTG AAG AAT GAT ATC ATA CCA TTC GTC ACA CTC TAT	360
101	Tyr Asn Arg Leu Val Asp Glu Leu Leu Lys Asn Asp Ile Ile Pro Phe Val Thr Leu Tyr	120
161	CAC TGG GAC TTA CCC TAC GCA CTT TAT GAA AAA GGT GGA TGG CTT AAC CCA GAT ATA GCG	420
121	His Trp Asp Leu Pro Tyr Ala Leu Tyr Glu Lys Gly Trp Leu Asn Pro Asp Ile Ala	140
421	CTC TAT TTC AGA GCA TAC GCA ACC TTT ATC TTC AAC GAA CTC GGT GAT CGT GTG AAA CAT	480
141	Leu Tyr Phe Arg Ala Tyr Ala Thr Phe Met Phe Asn Glu Leu Gly Asp Arg Val Lys His	160
481	TGG ATT ACA CTG AAC GAA CCA TGG TGT TCT TCT TCG GGT TAT TAC ACG GGA GAG CAT	540
161	Trp Ile Thr Leu Asn Glu Pro Trp Cys Ser Ser Phe Ser Gly Tyr Tyr Glu His	180
541	GCC CCG GGT CAT CAA AAT TTA CAA GAA GCG ATA ATC GCG GCG CAC AAC CTG TTG AGG GAA	600
181	Ala Pro Gly His Gln Asn Leu Gln Ala Ile Ile Ala His Asn Leu Leu Arg Glu	200
601	CAT GGA CAT GCC GTC CAG GCG TCC AGA GAA GAA GTC AAA GAT GGG GAA GTT GCC TTA ACC	660
201	His Gly His Ala Val Gln Ala Ser Arg Glu Glu Val Lys Asp Gly Glu Val Gly Leu Thr	220
661	AAC GTT GTG ATG AAA ATA GAA CCG GGC GAT GCA AAA CCC GAA AGT TTC TTG GTC GCA AGT	720
221	Asn Val Val Met Lys Ile Glu Pro Gly Asp Ala Lys Pro Glu Ser Phe Leu Val Ala Ser	240
721	CTT GTT GAT AAG TTC GTT AAT GCA TGG TCC CAT GAC CCT GTT GTT TTC GGA AAA TAT CCC	780
241	Leu Val Asp Lys Phe Val Asn Ala Trp Ser His Asp Pro Val Val Phe Gly Lys Tyr Pro	260
781	GAA GAA GCA GTT GCA CTT TAT ACG GAA AAA GGG TTG CAA GTT CTC GAT AGC GAT ATG AAT	840
261	Glu Glu Ala Val Ala Leu Tyr Thr Glu Lys Gly Leu Gln Val Leu Asp Ser Asp Met Asn	280
841	ATT ATT TCG ACT CCT ATA GAC TTC TTT GGT GTG AAT TAT TAC ACA AGA ACA CTT GTT GTT	900
281	Ile Ile Ser Thr Pro Ile Asp Phe Phe Gly Val Asn Tyr Tyr Arg Thr Leu Val Val	300
901	TTT GAT ATG AAC AAT CCT CTT GGA TTT TCG TAT GTT CAG GGA GAC CTT CCC AAA ACG GAG	960
301	Phe Asp Met Asn Asn Pro Leu Gly Phe Ser Tyr Val Gln Gly Asp Leu Pro Lys Thr Glu	320
961	ATG GGA TGG GAA ATC TAC CCG CAG GGA TTA TTT GAT ATG CTG GTC TAT CTG AAG GAA AGA	1020
321	Met Gly Trp Glu Ile Tyr Pro Gln Gly Leu Phe Asp Met Leu Val Tyr Leu Lys Glu Arg	340
1021	TAT AAA CTA CCA CTT TAT ATC ACA GAG AAC GGG ATG GCT GGA CCT GAT AAA TTG GAA AAC	1080
341	Tyr Lys Leu Pro Leu Tyr Ile Thr Glu Asn Gly Met Ala Gly Pro Asp Lys Leu Glu Asn	360
1081	GGA AGA GTT CAT GAT AAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT	1140
361	Gly Arg Val His Asp Asn Tyr Arg Ile Glu Tyr Leu Glu Lys His Phe Glu Lys Ala Leu	380
1141	GAA GCA ATC AAT GCA GAT GTT GAT TTG AAA GGT TAC TTC ATT TCG TCT TTG ATG GAT AAC	1200
381	Glu Ala Ile Asn Ala Asp Val Asp Leu Lys Gly Tyr Phe Ile Trp Ser Leu Met Asp Asn	400
1201	TTC GAA TGG GCG TCC GGA TAC TCC AAA CGT TTC GGT ATA ATC TAC GTC GAT TAC AAT ACC	1260
401	Phe Glu Trp Ala Cys Gly Tyr Ser Lys Arg Phe Gly Ile Ile Tyr Val Asp Tyr Asn Thr	420
1261	CCA AAA AGG ATA TTT AAA GAT TCA GCG ATG TCG TTG AAG GAA TTT CTA AAA TCT TAA	1317
421	Pro Lys Arg Ile Leu Lys Asp Ser Ala Met Trp Leu Lys Glu Phe Leu Lys Ser End	419

Figure 2

STAPHYLOTHERMUS MARINUS GLYCOSIDASE - 12G
 COMPLETE GENE SEQUENCE
 9/95

1	TTG ATA AGG TTT CCT GAT TAT TTC TTT GCA ATG CCT AGA TCA TCG CAA GAG ATG GAG	60
1	Met Ile Arg Phe Pro Asp Tyr Phe Leu Phe Gly Thr Ala Thr Ser Ser His Gln Ile Glu	60
61	GGT AAT AAC ATA TTT AAT GAT TCG TCG GAG TCG GAG ACT AAA GGC AGG ATT AAG CTT AGA	120
21	Gly Asn Asn Ile Phe Asn Asp Trp Trp Glu Trp Glu Thr Lys Gly Arg Ile Lys Val Arg	10
121	TCG CGT AAG GCA TGT AAT CAT TGG GAA CTC TAT AAA GAA GAC ATA GAG CTT ATG CCT GAG	180
41	Ser Gly Lys Ala Cys Asn His Trp Glu Leu Tyr Lys Glu Asp Ile Glu Leu Met Ala Glu	60
181	CTG GGA TAT AAT GCT TAT AGG TTC TCC ATA GAG TGG AGT AGA ATA TTT CCC AGA AAA GAT	240
61	Leu Gly Tyr Asn Ala Tyr Arg Phe Ser Ile Glu Trp Ser Arg Ile Phe Pro Arg Lys Asp	80
241	CAT ATA GAT TAT GAG TCG CTT AAT AAC TAT AAG GAA ATA GTT AAT CTA CTT AGA AAA TAC	300
81	His Ile Asp Tyr Glu Ser Leu Asn Lys Tyr Lys Glu Ile Val Asn Leu Leu Arg Lys Tyr	100
301	GGG ATA GAA CCT GTA ATC ACT CTT CAC CAC TTC ACA AAC CCG CAA TGG TTT ATG AAA ATT	360
101	Gly Ile Glu Pro Val Ile Thr Leu His His Phe Thr Asn Pro Gln Trp Phe Met Lys Ile	120
361	GGT GGA TCG ACT AGG GAA GAG AAC ATA AAA TAT TTT ATA AAA TAT GTA GAA CTT ATA GCT	420
121	Gly Gly Trp Thr Arg Glu Asn Ile Lys Tyr Phe Ile Lys Tyr Val Glu Leu Ile Ala	140
421	TCC GAG ATA AAA GAC GTG AAA ATA TGG ATC ACT ATT AAT GAA CCA ATA ATA TAT GTT TTA	480
141	Ser Glu Ile Lys Asp Val Lys Ile Trp Ile Thr Ile Asn Glu Pro Ile Ile Tyr Val Leu	160
481	CAA GGA TAT ATT TCC GCC GAA TGG CCA CCT GGA ATT AAA AAT TTA AAA ATA GCT GAT CAA	540
161	Gln Gly Tyr Ile Ser Gly Glu Trp Pro Pro Gly Ile Lys Asn Leu Lys Ile Ala Asp Gln	180
541	GTA ACT AAG AAT CTT TTA AAA GCA CAT AAT GAA GCC TAT AAT ATA CTT CAT AAA CAC GGT	600
181	Val Thr Lys Asn Leu Leu Lys Ala His Asn Glu Ala Tyr Asn Ile Leu His Lys His Gly	200
601	ATT GTA GCC ATA GCT AAA AAC ATG ATA GCA TTT AAA CCA GGA TCT AAT AGA GGA AAA GAC	660
201	Ile Val Gly Ile Ala Lys Asn Met Ile Ala Phe Lys Pro Gly Ser Asn Arg Gly Lys Asp	220
661	ATT AAT ATT TAT CAT AAA GTC GAT AAA GCA TTC AAC TGG GGA TTT CTC AAC GGA ATA TTA	720
221	Ile Asn Ile Tyr His Lys Val Asp Lys Ala Phe Asn Trp Gly Phe Leu Asn Gly Ile Leu	240
721	AGG GGA GAA CTA GAA ACT CTC CGT GGA AAA TAC CGA GTT GAG CCC GGA AAT ATT GAT TTC	780
241	Arg Gly Glu Leu Glu Thr Leu Arg Gly Lys Tyr Arg Val Glu Pro Gly Asn Ile Asp Phe	260
781	ATA GCC ATA AAC TAT TAT TCA TCA TAT ATT GTA AAA TAT ACT TGG AAT CCT TTT AAA CTA	840
261	Ile Gly Ile Asn Tyr Tyr Ser Ser Tyr Ile Val Lys Tyr Thr Trp Asn Pro Phe Lys Leu	280
841	CAT ATT AAA GTC GAA CCA TTA GAT ACA GGT CTA TGG ACA ACT ATG GGT TAC TGC ATA TAT	900
281	His Ile Lys Val Glu Pro Leu Asp Thr Gly Leu Trp Thr Thr Met Gly Tyr Cys Ile Tyr	300
901	CCT AGA GGA ATA TAT GAA GTT GTA ATG AAA ACT CAT GAG AAA TAC GCA AAA GAA ATA ATC	960
301	Pro Arg Gly Ile Tyr Glu Val Val Met Lys Thr His Glu Lys Tyr Gly Lys Glu Ile Ile	320
961	ATT ACA GAG AAC GGT GTT GCA GTA GAA AAT GAT GAA TTA AGG ATT TTA TCC ATT ATC AGG	1020
321	Ile Thr Glu Asn Gly Val Ala Val Glu Asn Asp Glu Leu Arg Ile Leu Ser Ile Ile Arg	340
1021	CAC TTA CAA TAC TTA TAT AAA CCC ATG AAT GAA GCA GCA AAG GTG AAA GGA TAT TTC TAC	1080
341	His Leu Gln Tyr Leu Tyr Lys Ala Met Asn Glu Gly Ala Lys Val Lys Gly Tyr Phe Tyr	360
1081	TGG ACC TTC ATC GAT AAT TTT GAG TGG GAT AAA GCA TTT AAC CAA ACC TTC GCA CTA GTC	1140
361	Trp Ser Phe Met Asp Asn Phe Glu Trp Asp Lys Gly Phe Asn Gln Arg Phe Gly Leu Val	380
1141	GAA GTT GAT TAT AAG ACT TTT GAG AGA AAA CCT AGA AAA ACC GCA TAT CTA TAT AGT CAA	1200
381	Glu Val Asp Tyr Lys Thr Phe Glu Arg Lys Pro Arg Lys Ser Ala Tyr Val Tyr Ser Gln	400
1201	ATA GCA CGT ACC AAG ACT ATA ACT GAT GAA TAC CTA GAA AAA TAT GCA TTA AAC AAC CTC	1260
401	Ile Ala Arg Thr Lys Thr Ile Ser Asp Glu Tyr Leu Glu Lys Tyr Gly Leu Lys Asn Leu	420
1261	GAA TAA 1266	
421	Glu End 422	

Figure 3

Thermococcus 9N1 Glycosidase -318/0
Complete gene sequence 9/95

1	ATG CTA CCA GAA GGC TTT CTC TGG GGC GTG TCC GAG TCC GGC TTT CAG TTC GAG ATG GGC	60
1	Met Leu Pro Glu Gly Phe Leu Trp Gly Val Ser Gln Ser Gly Phe Gln Phe Glu Met Gly	20
61	GAC AAG CTC ACG AGG AAC ATT GAT CCG AAC ACA GAC TGG TGG AAG TGG GTC AGG GAT CCC	120
21	Asp Lys Leu Arg Arg Asn Ile Asp Pro Asn Thr Asp Trp Trp Lys Trp Val Arg Asp Pro	40
121	TTC AAC ATA AAG AGG GAA CTC CTC AGC GGC GAC CTC CCC GAG GAG GGG ATA AAC AAC TAC	180
41	Phe Asn Ile Lys Arg Glu Leu Val Ser Gly Asp Leu Pro Glu Glu Gly Ile Asn Asn Tyr	60
181	GAA CTT TAC GAG AAG GAT CAC CCG CTC GGC AGA GAC CTC GGT CTC AAC GTC ATT TAC AGG ATT	140
61	Glu Leu Tyr Glu Lys Asp His Arg Leu Ala Arg Asp Leu Gly Leu Asn Val Tyr Arg Ile	80
241	CGA ATA GAG TGG AGG AGG ATC TTT CTC TGG GCA ACG TGG TTT GTG GAG GTT GAC GTT GAG	300
81	Gly Ile Glu Trp Ser Arg Ile Phe Pro Trp Pro Trp Phe Val Glu Val Asp Val Glu	100
301	CGG GAC AGC TAC GGA CTC GTG AAG GAC GTC AAA ATC GAT AAA GAC ACG CTC GAA GAG CTC	360
101	Arg Asp Ser Tyr Glu Leu Val Lys Asp Val Lys Ile Asp Lys Asp Thr Leu Glu Glu Leu	120
361	GAC GAG ATA GCG AAT CAT CAG GAG ATA GCC TAC TAC CGC CGC GTT ATA GAG AAC CTC AGC	420
121	Asp Glu Ile Ala Asn His Gln Glu Ile Ala Tyr Tyr Arg Arg Val Ile Glu His Leu Arg	140
421	GAG CTC GGC TTC AAG GTC ATC CTC AAC CAC TTC ACC CCC CTC TCG CTT CAC	480
141	Glu Leu Glu Phe Lys Val Ile Val Asn Leu Asn His Phe Thr Leu Pro Leu Trp Leu His	160
481	GAT CCC ATA ATC GCG ACG GAG AAC GGT CTC ACC AAC GGT AGC ATT GGC TCG GTC CGG CAG	540
161	Asp Pro Ile Ala Arg Glu Lys Ala Leu Thr Asn Gly Arg Ile Gly Trp Val Gly Gln	180
541	GAG ACC GTC GTG GAC TTC CCC AAG TAC GCG GCG TAC ATC GCG AAC GCA CTC GGG GAC CTC	600
181	Glu Ser Val Val Glu Phe Ala Lys Tyr Ala Ala Tyr Ile Ala Asn Ala Leu Gly Asp Leu	200
501	CTT CAT ATG TGG AGC ACC TTC AAC GAG CCG ATG GTC GTT GTG GAG CTC GGT TAC CTC GCT	660
201	Val Asp Met Trp Ser Thr Phe Asn Glu Pro Met Val Val Glu Leu Gly Tyr Leu Ala	220
661	CCC TAC TCC GGC TTT CGG CGG GGG GTT ATG AAC CCC GAG GCG GCA AAG CTC GCA ATC CTC	720
221	Pro Tyr Ser Glu Phe Pro Gly Val Met Asp Pro Glu Ala Ala Lys Leu Ala Ile Leu	240
721	AAC ATG ATA AAC GCC CAC GCA CTC CCC TAC AAG ATG ATA AAG AAG TTC GAC AGG GTC AAG	780
241	Asn Met Ile Asn Ala His Ala Leu Ala Tyr Lys Met Ile Lys Phe Asp Arg Val Lys	260
781	GCC GAT AAG GAT TCC CTC TCC GAG GGC GAG GTC GGC ATA ATC TAC AAC AAC ATA GGC GTT	840
261	Ala Asp Lys Asp Ser Arg Ser Glu Ala Glu Val Gly Ile Ile Tyr Asn Asn Ile Gly Val	280
841	GCC TAT CCA TAC GAC TCC AAC GAC CCA AAG GAC GTC AAA GCT GCA GAA AAC GAC AAC TAC	900
281	Ala Tyr Pro Tyr Asp Ser Asn Asp Pro Lys Asp Val Lys Ala Ala Glu Asn Asp Asn Tyr	300
901	TTC CAC ACC GGG CTC TTC TTC GAC GCA ATC CAC AAG GGC AAG CTC AAC ATC GAG TTC GAC	960
301	Phe His Ser Gly Leu Phe Phe Asp Ala Ile His Lys Gly Lys Leu Asn Ile Glu Phe Asp	320
961	GCT GAC ACC TTC GTC AAA GTT CGG CAT CTC AGC GGC AAC GAC TCG ATA GGC GTT AAC TAC	1020
321	Gly Glu Thr Phe Val Lys Val Arg His Leu Arg Gly Asn Asp Trp Ile Gly Val Asn Tyr	340
1021	TAC ACG AGA GAA GTC GTC AGG TAT TCG GAG CCC AAG TTC CCG AGC ATA CCC CTG ATA TCC	1080
341	Tyr Thr Arg Glu Val Val Arg Tyr Ser Glu Pro Lys Phe Pro Ser Ile Pro Leu Ile Ser	360
1081	TTC CGG GCA CTT CAC AAC TAC GGC TAC GGC TCC AGG CCC GCG ACT TCT TCC GGC GAC GGA	1140
361	Phe Arg Gly Val His Asn Tyr Gly Tyr Ala Cys Arg Pro Gly Ser Ser Ser Ala Asp Gly	380
1141	AGG CCC GTC AGC GAC ATC GGC TCG GAG ATC TAT CCG GAG GGG ATC TAC GAC TCG ATA AGA	1200
381	Arg Pro Val Ser Asp Ile Glu Trp Glu Ile Tyr Pro Glu Gly Ile Tyr Asp Ser Ile Arg	400
1201	GAG GCC AAC AAA TAC GGG GTC CCC CTT TAC GTC ACC GAA AAC GGA ATA GCC GAT TCA ACT	1260
401	Glu Ala Asn Lys Tyr Glu Val Pro Val Tyr Val Thr Glu Asn Gly Ile Ala Asp Ser Thr	420
1261	GAC ACC CTG CGG CGG TAC CTC GCG ACC CAT GTC CGG AAG ATT CGAG GAG GCG TAC GAG	1320
421	Asp Thr Leu Arg Pro Tyr Tyr Leu Ala Ser His Val Ala Lys Ile Glu Glu Ala Tyr Glu	440

Figure 4

1321	CCG CGT TAC GAC GTC ACC GGC TAC CTT TAC TGG GCG CTC ACC GAC AAC TAC GAG TCG GCC	1380
441	Ala Gly Tyr Asp Val Arg Gly Tyr Leu Tyr Trp Ala Leu Thr Asp Asn Tyr Glu Trp Ala	460
1381	CTC CGT TTC AGG ATG ACC TTC GGC CTC TAT AAA GIG GAT CTC ATA ACC AAG GAG AGA ACA	1440
461	Leu Gly Phe Arg Met Arg Phe Gly Leu Tyr Lys Val Asp Leu Ile Thr Lys Glu Arg Thr	480
1441	CCG CGG GAG GAA ACC GTC AAG CTT TAT ACC CCC ATC CTC GAG AAC AAC GGA GTC AGC AAC	1500
481	Pro Arg Glu Glu Ser Val Lys Val Tyr Arg Gly Ile Val Glu Asp Asn Gly Val Ser Lys	500
1501	GAA ATC CGG GAG AAG TTC GCA CTT GGG TCA	1530
501	Glu Ile Arg Glu Lys Phe Gly Leu Gly End	510

Figure 4 (Continued)

1	ATG	GAA	AGG	ATC	GAT	GAA	ATT	CTC	TCT	GAG	TTA	ACT	ACA	GAG	GAA	AAG	CTG	AAG	CTC	CTT	AT	N
1	Met	Glu	Arg	Ile	Asp	Glu	Ile	Leu	Ser	Glu	Ile	Thr	Thr	Glu	Glu	Lys	Val	Ile	Ile	Leu	Val	20
61	GTG	GCG	GTT	GCT	CTT	CCA	GGA	CTT	TTT	GCG	AAC	CCA	CAT	TCC	AGA	CTG	GCG	GCT	GCG	GCT	AT	120
21	Val	Gly	Val	Gly	Ile	Pro	Gly	Leu	Phe	Gly	Asn	Pro	His	Ser	Arg	Val	Ala	Gly	Ala	Ala	Ala	40
121	GGA	GAA	ACA	CAT	CCC	CTT	CCA	AGA	CTT	GGA	ATT	CCT	GCG	TTT	GTC	CTG	GCA	GAT	GGT	CCC	180	
41	Gly	Glu	Thr	Ile	Pro	Val	Pro	Arg	Ile	Gly	Ile	Pro	Ala	Phe	Val	Ile	Ala	Gly	Pro	Ala	60	
181	GCA	GGA	CTC	AGA	ATA	AAT	CCC	ACA	AGG	GAA	AAC	GAT	GAA	AAC	ACT	TAC	TAC	ACG	ACG	GCA	240	
61	Ala	Gly	Ile	Leu	Arg	Ile	Asn	Pro	Thr	Arg	Glu	Asn	Asp	Glu	Asn	Thr	Tyr	Thr	Thr	Ala	80	
241	TTT	CCC	GTT	GAA	ATC	ATG	CTC	GCT	TCT	ACC	TGG	AAC	AGA	GAC	CTT	CTG	GAA	GAA	GTG	GGA	300	
81	Phe	Pro	Val	Gly	Ile	Met	Leu	Ala	Ser	Thr	Trp	Asn	Arg	Asp	Leu	Leu	Glu	Glu	Val	Gly	100	
301	AAA	GCC	ATG	GGA	GAA	GAA	GTT	AGG	GAA	TAC	GGT	GTC	GAT	GTG	CTT	CTT	GCA	CCT	GCG	ATG	360	
101	Lys	Ala	Met	Gly	Glu	Glu	Val	Arg	Glu	Tyr	Gly	Val	Asp	Val	Leu	Leu	Ala	Pro	Ala	Met	120	
361	AAC	ATT	CAC	AGA	AAC	CCT	CTT	TGT	GGA	AGG												

Figure 5

1201	GAG	GAG	TAC	ATA	AAA	AAG	ATG	AGA	GAA	ACA	GAG	GAA	TAT	AAA	CCC	AGA	ACC	GAC	TCT	TGG	1200
401	Glu	Glu	Tyr	Ile	Lys	Lys	Met	Arg	Glu	Thr	Glu	Glu	Tyr	Lys	Pro	Arg	Thr	Asp	Ser	Trp	420
1261	GGA	ACG	GTC	ATA	AAA	CCG	AAA	CTC	CCA	GAG	AAT	TTC	CTC	TCA	GAA	AAA	GAG	ATA	AAG	AAA	1320
421	Gly	Thr	Val	Ile	Lys	Pro	Lys	Lys	Glu	Asn	Phe	Leu	Ser	Glu	Glu	Lys	Glu	Ile	Lys	440	
1321	CCT	CCA	AAG	AAA	AAC	GAT	GTT	GCA	GTT	GTT	GTG	ATC	ACT	AGG	ATC	TCC	GCT	GAG	GGA	TAC	1380
441	Pro	Pro	Lys	Lys	Asn	Asp	Val	Ala	Val	Val	Val	Ile	Ser	Arg	Ile	Ser	Gly	Glu	Gly	Tyr	460
1381	GAC	AGA	AAG	CCG	GTC	AAA	GGT	GAC	TTC	TAC	CTC	TCC	GAT	GAC	GAG	CTG	GAA	CTC	ATA	AAA	1440
461	Asp	Arg	Lys	Pro	Val	Lys	Gly	Asp	Phe	Tyr	Leu	Ser	Asp	Asp	Glu	Leu	Glu	Leu	Ile	Lys	480
1441	ACC	GTC	TCG	AAA	GAA	TTC	CAC	GAT	CAG	GGT	AAG	AAA	GTT	GTC	GTT	CTT	CTG	AAC	ATC	GGA	1500
481	Thr	Val	Ser	Lys	Gly	Phe	His	Asp	Gln	Gly	Lys	Lys	Val	Val	Val	Leu	Leu	Asn	Ile	Gly	500
1501	AGT	CCC	ATC	GAA	GTC	GCA	AGC	TGG	AGA	GAC	CTT	GTC	GAT	GGA	ATT	CTT	CTC	GTC	TGG	CAG	1560
501	Ser	Pro	Ile	Glu	Val	Ala	Ser	Trp	Arg	Asp	Leu	Val	Asp	Gly	Ile	Leu	Leu	Val	Trp	Gln	520
1561	GCG	GGA	CAG	GAG	ATG	GGA	AGA	ATA	GTC	GCC	GAT	GTT	CTT	GTG	GGA	AAG	ATT	AAT	CCC	TCC	1620
521	Ala	Gly	Glu	Glu	Met	Gly	Arg	Ile	Val	Ala	Asp	Val	Gly	Lys	Lys	Ile	Asn	Pro	Ser	540	
1621	GGA	AAA	CTT	CCA	ACG	ACC	TTC	CCG	AAG	GAT	TAC	TCG	GAC	GTT	CCA	TCC	TGG	ACG	TTC	CCA	1680
541	Gly	Lys	Leu	Pro	Tyr	Thr	Phe	Pro	Lys	Asp	Tyr	Ser	Asp	Val	Pro	Ser	Trp	Thr	Phe	Pro	560
1681	GGA	GAG	CCA	AAG	GAC	AAT	CCG	CAA	AGA	GTC	GTC	TAC	GAG	GAA	GAC	ATC	TAC	GTG	GGA	TAC	1740
561	Gly	Glu	Pro	Lys	Asp	Asn	Pro	Gln	Arg	Val	Val	Tyr	Glu	Glu	Asp	Ile	Tyr	Val	Gly	Tyr	580
1741	AGG	TAC	TAC	GAC	ACC	TTC	GGT	GTG	GAA	CCT	GCC	TAC	GAA	TTC	GCC	TAC	GGC	CTC	TCT	TAC	1800
581	Arg	Tyr	Tyr	Asp	Trp	Phe	Gly	Val	Glu	Pro	Ala	Tyr	Glu	Phe	Gly	Tyr	Gly	Ile	Ser	Tyr	600
1801	ACA	AAG	TTT	GAA	TAC	AAA	GAT	TTA	AAA	ATC	GCT	ATC	GAC	GGT	GAG	ACG	CTC	AGA	GTG	TCG	1860
601	Thr	Lys	Phe	Glu	Tyr	Lys	Asp	Leu	Lys	Ile	Ala	Ile	Asp	Gly	Glu	Thr	Leu	Arg	Val	Ser	620
1861	TAC	ACG	ATC	ACA	AAC	ACT	GGG	GAC	AGA	GCT	GGA										

Figure 5 (Continued)

THERMOCOCCUS AEDII12RA GLYCOSIDASE (18B/G)
COMPLETE GENE SEQUENCE - 9/95

1	ATG ATC CAC TCC CCC GTT AAA CGG ATT ATA TCT GAG GCT CCC GCC ATA ACC ATC ACA ATA	60
1	Met Ile His Cys Pro Val Lys Gly Ile Ile Ser Glu Ala Arg Gly Ile Thr Ile Thr Ile	20
61	GAT TTA AGT TTT CAA GGC CAA ATA AAT AAT TTG CTG AAT GCT ATC ATT GTC TTT CCG GAG	120
21	Asp Leu Ser Phe Gln Gly Ile Asn Asn Leu Val Asn Ala Met Ile Val Phe Pro Glu	40
121	TTC TPC CTC TTT GGA ACC GCC ACA TCT TCT CAT CAG ATC GAG GGA GAT AAT AAA TGG AAC	180
41	Phe Phe Leu Phe Gly Thr Ala Thr Ser Ser His Gln Ile Glu Gly Asp Asn Lys Trp Asn	60
181	GAC TGG TCG TAT TAT GAG GAG ATA GGT AAG CTC CCC TAC AAA TCC GGT AAA GCC TGC AAT	240
61	Asp Trp Trp Tyr Tyr Glu Glu Ile Gly Lys Leu Pro Tyr Lys Ser Gly Lys Ala Cys Asn	80
241	CAC TGG GAG CTT TAC AGG GAA GAT ATA GAG CTA ATG GCA CAG CTC GCC TAC AAT GCC TAC	300
81	His Trp Glu Leu Tyr Arg Glu Asp Ile Glu Leu Met Ala Gln Leu Gly Tyr Asn Ala Tyr	100
301	CGC TTT TCG ATA GAG TGG ACC CGT CTC TTC CCG GAA GAG GCC AAA TTC AAT GAA GAA GCC	360
101	Arg Phe Ser Ile Glu Trp Ser Arg Leu Phe Pro Glu Glu Gly Lys Phe Asn Glu Glu Ala	120
361	TTC AAC CGC TAC CGT GAA ATA ATT GAA ATC CTC CTT GAG AAG GGG ATT ACT CCA AAC GTT	420
121	Phe Asn Arg Tyr Arg Glu Ile Ile Glu Leu Leu Lys Gly Ile Thr Pro Asn Val	140
421	ACA CTG CAC CAC TTC ACA TCA CCG CTG TGG TTC ATG CGG AAG GGA GGC TTT TTG AAG GAA	480
141	Thr Leu His His Phe Thr Ser Pro Leu Trp Phe Met Arg Lys Gly Gly Phe Leu Lys Glu	160
481	GAA AAC CTC AAG TAC TGG GAG CAG TAC GTT GAT AAA GCC GCG GAG CTC CTC AAG GGA GTC	540
161	Glu Asn Leu Lys Tyr Trp Glu Gln Tyr Val Asp Lys Ala Ala Glu Leu Leu Lys Gly Val	180
541	AAG CTT GTA GCT ACA TTC AAC GAG CCG ATG GTC TAT GTT ATG ATG GGC TAC CTC ACA GCC	600
181	Lys Leu Val Ala Thr Phe Asn Glu Pro Met Val Tyr Val Met Met Gly Tyr Leu Thr Ala	200
601	TAC TGG CCG CCC TTC ATC AAG AGT CCC TTT AAA GCC TTT AAA GTT GCC GCA AAC CTC CTT	660
201	Tyr Trp Pro Pro Phe Ile Lys Ser Pro Phe Lys Ala Phe Lys Val Ala Ala Asn Leu Leu	220
661	AAG GCC CAT GCA ATG GCA TAT GAT ATC CTC CAT GCT AAC TTT GAT GTG GGG ATA GTT AAA	720
221	Lys Ala His Ala Met Ala Tyr Asp Ile Leu His Gly Asn Phe Asp Val Gly Ile Val Lys	240
721	AAC ATC CCC ATA ATG CTC CCT GCA AGC AAC AGA GAG AAA GAC GTA GAA GCT GCC CAA AAG	780
241	Asn Ile Pro Ile Met Leu Pro Ala Ser Asn Arg Glu Lys Asp Val Glu Ala Ala Gln Lys	260
781	GCG GAT AAC CTC TTT AAC TGG AAC TTC CTT GAT GCA ATA TGG ACC GGA AAA TAT AAA GGA	840
261	Ala Asp Asn Leu Phe Asn Trp Asn Phe Leu Asp Ala Ile Trp Ser Gly Lys Tyr Lys Gly	280
841	GCT TTT GGA ACT TAC AAA ACT CCA GAA AGC GAT GCA GAC TTC ATA GGG ATA AAC TAC TAC	900
281	Ala Phe Gly Thr Tyr Lys Thr Pro Glu Ser Asp Ala Asp Ile Gly Ile Asn Tyr Tyr	300
901	ACA GCC AGC GAG GTA AGG CAT AGC TGG AAT CCG CTA AAG TTT TTC TAC GAT GCC AAG CTT	960
301	Thr Ala Ser Glu Val Arg His Ser Trp Asn Pro Leu Lys Phe Phe Asp Ala Lys Leu	320
961	GCA GAC TTA AGC GAG AGA AAA ACA GAT ATG GGT TGG AGT GTC TAT CCA AAG GGC ATA TAC	1020
321	Ala Asp Leu Ser Glu Arg Lys Thr Asp Met Gly Trp Ser Val Tyr Pro Lys Gly Ile Tyr	340
1021	GAA GCT ATA GCA AAG GTT TCA CAC TAC GGA AAG CCA ATG TAC ATC ACG GAA AAC GGG ATA	1080
341	Glu Ala Ile Ala Lys Val Ser His Tyr Gly Lys Pro Met Tyr Ile Thr Glu Asn Gly Ile	360
1081	GCT ACC TTA GAC GAT GAG TGG AGG ATA GAG TTT ATC ATC CAG CAC CTC CAG TAC GTT CAC	1140
361	Ala Thr Leu Asp Asp Glu Trp Arg Ile Glu Phe Ile Ile Gln His Leu Gln Tyr Val His	380
1141	AAA GCC TTA AAC GAT GGC TTT GAC TTG AGA GGC TAC TTC TAT TGG TCT TTT ATG GAT AAC	1200
381	Lys Ala Leu Asn Asp Gly Phe Asp Leu Arg Gly Tyr Phe Tyr Ser Phe Met Asp Asn	400
1201	TTC GAG TGG GCT GAG GGT TTT AGA CCA CGC TTT GGG CTC GTC GAG GTC GAC TAC ACG ACC	1260
401	Phe Glu Trp Ala Glu Gly Phe Arg Pro Arg Phe Gly Leu Val Glu Val Asp Tyr Thr Thr	420
1261	TTC AAG AGG AGA CCG AGA AAG AGT GCT TAC ATA TAT GGA GAA ATT CCA AGG GAA AAG AAA	1320
421	Phe Lys Arg Arg Pro Arg Lys Ser Ala Tyr Ile Tyr Gly Glu Ile Ala Arg Glu Lys Lys	440
1321	ATA AAA GAC GAA CTG CTG GCA AAG TAT GGG CTT CCG GAG CTA TGA 1365	
441	Ile Lys Asp Glu Leu Leu Ala Lys Tyr Gly Leu Pro Glu Leu End 455	

Figure 6

WO 97/25417

THERMOCOCCUS CHITONOPHAGUS GLYCOSIDASE - 22G
COMPLETE SEQUENCE - 9/95

1	TTG CTT CCA GAG AAC TTT CTC TCG GGA GTT TCA CAG TCC GGA TTC CAC TTT GAA ATC GGG	60
1	Met Leu Pro Glu Asn Phe Leu Trp Gly Val Ser Gln Ser Gly Phe Gln Phe Glu Met Gly	20
61	GAC AGA CTG AGG AGG CAC ATT GAT CCA AAC ACA GAT TCG TGG TAC TCG GTC AGA GAT GAA	120
21	Asp Arg Leu Arg Arg His Ile Asp Pro Asn Thr Asp Trp Trp Tyr Trp Val Arg Asp Glu	40
121	TAT AAT ATC AAA AAA GGA CTA GTC AGT GGG GAT CTT CCC GAA GAC GGT ATA AAT TCA TAT	180
41	Tyr Asn Ile Lys Lys Gly Leu Val Ser Gly Asp Leu Pro Glu Asp Gly Ile Asn Ser Tyr	60
181	GAA TTA TAT GAG AGA GAC CAA GAA ATT GCA AAG GAT TTA GGG CTC AAC ACA TAT AGG ATC	240
61	Glu Leu Tyr Glu Arg Asp Gln Glu Ile Ala Lys Asp Leu Gly Leu Asn Thr Tyr Arg Ile	80
241	GGA ATT CAA TGG AGC AGA GTC TTT CCA TGG CCA ACG ACT TTT GTC GAC GTG GAG TAT GAA	300
81	Gly Ile Glu Trp Ser Arg Val Phe Pro Trp Pro Thr Phe Val Asp Val Glu Tyr Glu	100
301	ATT GAT GAG TCT TAC CGG TTG GTC AAG GAT GTG AAG ATT TCT AAA GAC GCA TTA GAA AAA	360
101	Ile Asp Glu Ser Tyr Gly Leu Val Lys Asp Val Lys Ile Ser Lys Asp Ala Leu Glu Lys	120
361	CTT GAT GAA ATC GCT AAC CAA ACG GAA ATA ATA TAT TAT AGG AAC CTA ATA AAT TCC CTA	420
121	Leu Asp Glu Ile Ala Asn Gln Arg Glu Ile Ile Tyr Tyr Arg Asn Leu Ile Asn Ser Leu	140
421	AGA AAG AGG GGT TTT AAG GTC ATA CTA AAC CTA AAT CAT TTT ACC CTC CCA ATA TGG CTT	480
141	Arg Lys Arg Gly Phe Lys Val Ile Leu Asn Leu Asn His Phe Thr Leu Pro Ile Trp Leu	160
481	CAT GAT CCT ATC GAA TCT AGA GAA AAA GCC CTG ACC AAT AAG AGA AAC GGA TGG GTC AGC	540
161	His Asp Pro Ile Glu Ser Arg Glu Lys Ala Leu Thr Asn Lys Arg Asn Gly Trp Val Ser	180
541	GAA AGG AGT GTT ATA GAG TTT GCA AAA TTT GCC GCG TAT TTA GCA TAT AAA TTC GGA GAC	600
181	Glu Arg Ser Val Ile Glu Phe Ala Lys Phe Ala Ala Tyr Leu Ala Tyr Lys Phe Gly Asp	200
601	ATA GTC GAC ATG TGG AGC ACA TTT AAT GAA CCT ATG GTG GTC GCC GAG TTG GGG TAT TTA	660
201	Ile Val Asp Met Trp Ser Thr Phe Asn Glu Pro Met Val Val Ala Glu Leu Gly Tyr Leu	220
661	GCC CCA TAC TCA GGA TTC CCC CCG GGA GTC ATG AAT CCA GAA GCA GCA AAG TTA GTT ATG	720
221	Ala Pro Tyr Ser Gly Phe Pro Pro Gly Val Met Asn Pro Glu Ala Ala Lys Leu Val Met	240
721	CTA CAT ATG ATA AAC GCC CAT GCT TTA GCA TAT AGG ATG ATA AAG AAA TTT GAC AGA AAA	780
241	Leu His Met Ile Asn Ala His Ala Leu Ala Tyr Arg Met Ile Lys Lys Phe Asp Arg Lys	260
781	AAA GCT GAT CCA GAA TCA AAA GAA CCA GCT GAA ATA GGA ATT ATA TAC AAT AAC ATC GGC	840
261	Lys Ala Asp Pro Glu Ser Lys Glu Pro Ala Glu Ile Gly Ile Ile Tyr Asn Asn Ile Gly	280
841	GTC ACA TAT CCG TTT AAT CCG AAA GAC TCA AAG GAT CTA CAA GCA TCC GAT AAT GCC AAT	900
281	Val Thr Tyr Pro Phe Asn Pro Lys Asp Ser Lys Asp Leu Gln Ala Ser Asp Asn Ala Asn	300
901	TTC TTC CAC AGT GGG CTA TTC TTA ACG GCT ATC CAC AGG GGA AAA TTA AAT ATC GAA TTT	960
301	Phe Phe His Ser Gly Leu Phe Leu Thr Ala Ile His Arg Gly Lys Leu Asn Ile Glu Phe	320
961	GAC GGA GAG ACA TTT GTT TAC CTT CCA TAT TTA AAG GGC AAT GAT TGG CTG GGA GTG AAT	1020
321	Asp Gly Glu Thr Phe Val Tyr Leu Pro Tyr Leu Lys Gly Asn Asp Trp Leu Gly Val Asn	340
1021	TAT TAT ACA AGA GAA GTC GTT AAA TAC CAA GAT CCC ATG TTT CCA AGT ATC CCT CTC ATA	1080
341	Tyr Tyr Thr Arg Glu Val Val Lys Tyr Gln Asp Pro Met Phe Pro Ser Ile Pro Leu Ile	360
1081	AGC TTC AAG GGC GTT CCA GAT TAT GGA TAC GGA TCT AGA CCA GGA ACG ACG TCA AAG GAC	1140
361	Ser Phe Lys Gly Val Pro Asp Tyr Gly Tyr Gly Cys Arg Pro Gly Thr Thr Ser Lys Asp	380
1141	GGT AAT CCT GTT AGT GAC ATT GGA TCG GAG GTC TAT CCC AAA GGC ATG TAC GAC TCT ATA	1200
381	Gly Asn Pro Val Ser Asp Ile Glu Trp Glu Val Tyr Pro Lys Gly Met Tyr Asp Ser Ile	400
1201	GTC GCT GCC AAT GAA TAT GGA GTT CCT GTC TAC GTC ACA GAA AAC GGA ATA GCA GAT TCA	1260
401	Val Ala Ala Asn Glu Tyr Gly Val Pro Val Tyr Val Thr Glu Asn Gly Ile Ala Asp Ser	420
1261	AAA GAT GTC TTA AGG CCC TAT TAC ATC GCA TCT CAC ATT GAA GCC ATG GAA GAG GCT TAC	1320
421	Lys Asp Val Leu Arg Pro Tyr Tyr Ile Ala Ser His Ile Glu Ala Met Glu Glu Ala Tyr	440

Figure 7

1321	CAA AAT CCT TAT GAC GTC AGA GCA TAC TTA CAC TGG GCA TTA ACC GAT AAT TAC GAA TCG	1380
441	Glu Asn Gly Tyr Asp Val Arg Gly Tyr Leu His Trp Ala Leu Thr Asp Asn Tyr Glu Trp	460
1381	CCC TTA GGG TTC AGA ATG AGG TTT CCC TTG TAC GAA GCA AAC TTG ATA ACC AAA GAG AGA	1440
461	Ala Leu Gly Phe Arg Met Arg Phe Gly Leu Tyr Glu Val Asn Leu Ile Thr Lys Glu Arg	480
1441	AAA CCC AGG AAA AAG ACT GCA AGA GCA TTC AGA GAG ATA GTT ATT AAT AAT GGG CTA ACA	1500
481	Lys Pro Arg Lys Lys Ser Val Arg Val Phe Arg Glu Ile Val Ile Asn Asn Gly Leu Thr	500
1501	AGC AAC ATC AGG AAA GAG ATC TTA CAG GAG CGG TAG	1536
501	Ser Asn Ile Arg Lys Glu Ile Leu Glu Glu Gly End	512

Figure 7 (Continued)

PYROCOCCUS FURIOSUS GLYCOSIDASE - 7G1
COMPLETE GENE SEQUENCE - 10/95

1	ATG	TTC	CCT	GAA	AAG	TTC	CTT	TGG	GGT	GCA	CAA	TCG	GGT	TTT	CAG	TTT	GAA	ATG	GGG	60	
1	Met	Phe	Pro	Glu	Lys	Phe	Leu	Irp	Gly	Val	Ala	Gln	Ser	Gly	Phe	Gin	Phe	Glu	Met	Gly	20
61	GAT	AAA	CTC	AGC	AGG	AAT	ATT	GAC	ACT	AAC	ACT	GAT	TGG	TGG	CAC	TGG	GCA	AGC	GAT	AAG	120
21	Asp	Lys	Leu	Arg	Arg	Asn	Ile	Asp	Thr	Asn	Thr	Asp	Trp	Trp	His	Trp	Val	Arg	Asp	Lys	40
121	ACA	AAT	ATA	GAG	AAA	GCC	CTC	GTT	AGT	GGA	GAT	CTT	CCC	GAG	GAG	GGG	ATT	AAC	AAT	TAC	180
41	Thr	Asn	Ile	Glu	Lys	Gly	Leu	Val	Ser	Gly	Asp	Leu	Pro	Glu	Gly	Ile	Asn	Asn	Tyr	60	
181	GAG	CTT	TAT	GAC	AAG	GAC	CAT	GAG	ATT	GCA	AGA	AAG	CTG	GGT	CTT	AAT	GCT	TAC	AGA	ATA	240
61	Glu	Leu	Tyr	Glu	Lys	Asp	His	Glu	Ile	Ala	Arg	Lys	Leu	Gly	Leu	Asn	Ala	Tyr	Arg	Ile	80
241	GCC	ATA	GAG	TGG	AGC	AGA	ATA	TTC	CCA	TGG	CCA	ACG	ACA	TTT	ATT	GAT	GTT	GAT	TAT	AGC	300
81	Gly	Ile	Glu	Trp	Ser	Arg	Ile	Phe	Pro	Trp	Pro	Thr	Thr	Phe	Ile	Asp	Val	Asp	Tyr	Ser	100
301	TAT	AAT	GAA	TCA	TAT	AAC	CTT	ATA	GAA	GAT	GTA	AAG	ATC	ACC	AAG	GAC	ACT	TTG	GAG	GAG	360
102	Tyr	Asn	Glu	Ser	Tyr	Asn	Leu	Ile	Glu	Asp	Val	Lys	Ile	Thr	Lys	Asp	Thr	Leu	Glu	Ile	120
361	TTA	GAT	GAG	ATC	GCC	AAC	AAG	AGG	GAG	GTC	TAC	TAT	AGG	TCA	GTC	ATA	AAC	AGC	CTG	420	
121	Leu	Asp	Glu	Ile	Ala	Asn	Lys	Arg	Glu	Val	Ala	Tyr	Tyr	Ser	Val	Ile	Asn	Ser	Leu	140	
421	AGG	AGC	AAG	GGG	TTT	AAG	GTT	ATA	GTT	AAT	CTA	AAT	CAC	TTC	ACC	CTT	CCA	TAT	TGG	TTG	480
141	Arg	Ser	Lys	Gly	Phe	Lys	Val	Ile	Val	Asn	Leu	Asn	His	Phe	Thr	Leu	Pro	Tyr	Trp	Leu	160
481	CAT	GAT	CCC	ATT	GAG	GCT	AGG	GAG	GCG	TTA	ACT	AAT	AAG	AGG	AAC	GCG	TCG	GTT	AAC	540	
161	His	Asp	Pro	Ile	Glu	Ala	Arg	Glu	Arg	Ala	Leu	Thr	Asn	Lys	Arg	Asn	Gly	Trp	Val	Asn	180
541	CCA	AGA	ACA	GTC	ATA	GAG	TTT	GCA	AAG	TAT	GCC	GCT	TAC	ATA	GCC	TAT	AAG	TTT	GGA	GAT	600
181	Pro	Arg	Thr	Val	Ile	Glu	Phe	Ala	Lys	Tyr	Ala	Ala	Tyr	Ile	Ala	Tyr	Lys	Phe	Gly	Asp	200
601	ATA	GTG	GAT	ATG	TGG	AGC	ACG	TTT	AAT	GAG	CCT	ATG	GTG	GTT	GAG	CTT	GGC	TAC	CTA	660	
201	Ile	Val	Asp	Met	Trp	Ser	Thr	Phe	Asn	Glu	Pro	Met	Val	Val	Val	Glu	Leu	Gly	Tyr	Ile	220
661	GCC	CCC	TAC	TCT	GGC	TTC	CCT	CCA	GGG	GTT	CTA	AAT	CCA	GAG	GCC	GCA	AAG	CTG	GCG	ATA	720
221	Ala	Pro	Tyr	Ser	Gly	Phe	Pro	Pro	Gly	Val	Leu	Asn	Pro	Glu	Ala	Ala	Lys	Leu	Ala	Ile	240
721	CTT	CAC	ATG	ATA	AAT	GCA	CAT	GCT	TTA	GCT	TAT	AGG	CAG	ATA	AAG	AAG	TTT	GAC	ACT	GAG	780
241	Leu	His	Met	Ile	Asn	Ala	His	Ala	Leu	Ala	Tyr	Arg	Gln	Ile	Lys	Lys	Phe	Asp	Thr	Glu	260
781	AAA	GCT	GAT	AAG	GAT	TCT	AAA	GAG	CCT	GCA	GAA	GTT	ATA	ATT	TAC	AAC	AAC	ATT	GGG	840	
261	Lys	Ala	Asp	Lys	Asp	Ser	Lys	Glu	Pro	Ala	Glu	Val	Gly	Ile	Ile	Tyr	Asn	Asn	Ile	Gly	280
841	GTT	GCT	TAT	CCC	AAG	GAT	CCG	AAC	GAT	TCC	AAG	GAT	GTT	AAG	GCA	GCA	GAA	AAC	GAC	900	
261	Val	Ala	Tyr	Pro	Lys	Asp	Pro	Asn	Asp	Ser	Lys	Asp	Val	Lys	Ala	Ala	Glu	Asn	Asp	Asn	300
901	TTC	TTC	CAC	TCA	GGG	CTG	TTC	GAG	GCC	ATA	CAC	AAA	GGA	AAA	CTT	AAT	ATA	GAG	TTT	960	
301	Phe	Phe	His	Ser	Gly	Leu	Phe	Phe	Glu	Ala	Ile	His	Lys	Gly	Lys	Leu	Asn	Ile	Glu	Phe	320
961	GAC	GGT	GAA	ACG	TTT	ATA	GAT	GCC	CCC	TAT	CTA	AAG	GGC	AAT	GAC	TCG	ATA	GGG	GTT	1020	
321	Asp	Gly	Glu	Thr	Phe	Ile	Asp	Ala	Pro	Tyr	Leu	Lys	Gly	Asn	Asp	Trp	Ile	Gly	Val	Asn	340
1021	TAC	TAC	ACA	AGG	GAA	GTA	GTT	ACG	TAT	CAG	GAA	CCA	ATG	TTT	CCT	TCA	ATC	CCG	CTG	ATC	1080
341	Tyr	Tyr	Thr	Arg	Glu	Val	Val	Thr	Tyr	Gin	Glu	Pro	Met	Phe	Pro	Ser	Ile	Pro	Leu	Ile	360
1081	ACC	TTT	AAG	GGA	GTT	CAA	CGA	TAT	GCC	TAT	GCA	AGA	CCT	GGG	ACT	CTG	TCA	AAG	GAT	1140	
361	Thr	Phe	Lys	Gly	Val	Gln	Gly	Tyr	Gly	Tyr	Ala	Cys	Arg	Pro	Gly	Thr	Leu	Ser	Lys	Asp	380
1141	GAC	AGA	CCC	GTC	AGC	GAC	ATA	GGA	TGG	GAA	CTC	TAT	CCA	GAG	GGG	ATG	TAC	GAT	TCA	1200	
381	Asp	Arg	Pro	Val	Ser	Asp	Ile	Gly	Tyr	Glu	Leu	Tyr	Pro	Glu	Gly	Met	Tyr	Asp	Ser	Ile	400
1201	GTT	GAA	GCT	CAC	AAG	TAC	GGC	GTT	CCA	GTT	TAC	GTC	ACG	GAG	AAC	GGG	ATA	GCG	GAT	TCA	1260
401	Val	Glu	Ala	His	Lys	Tyr	Gly	Val	Pro	Val	Tyr	Val	Asn	Gly	Ile	Ala	Asp	Ser		420	

Figure 8

1261 AAG GAC ATC CTA AGA CCT TAC TAC ATA GCG AGC CAC ATA AAG ATG ATA GAG AAG GCC TTT 1320
421 Lys Asp Ile Leu Arg Pro Tyr Tyr Ile Ala Ser His Ile Lys Met Ile Glu Lys Ala Phe 440

1321 GAG GAT GGG TAT GAA GTC AAG GGC TAC TTC CAC TGG GCA TTA ACT GAC AAC TTC GAG TGG 1380
441 Glu Asp Gly Tyr Glu Val Lys Gly Tyr Phe His Trp Ala Leu Thr Asp Asn Phe Glu Trp 460

1381 GCT CTC GGG TTT AGA ATG CGC TTT GGC CTC TAC GAA GTC AAC CTA ATT ACA AAG GAG AGA 1440
461 Ala Leu Gly Phe Arg Met Arg Phe Gly Leu Tyr Glu Val Asn Leu Ile Thr Lys Glu Arg 480

1441 ATT CCC AGG GAG AAG AGC GTG ATA TTC AGA GAG ATA GTA GCC AAT AAT GGT GTT ACG 1500
481 Ile Pro Arg Glu Ser Val Ser Ile Phe Arg Glu Ile Val Ala Asn Asn Gly Val Thr 500

1501 AAA AAG ATT GAA GAG GAA TTG CTG AGG GGA TGA 1533
501 Lys Lys Ile Glu Glu Leu Leu Arg Gly End 511

Figure 8 (Continued)

Bankia gouldi endoglucanase (37GP1)

9 18 27 36 45 54
 5 ATG AGA ATA CGT TTA GCG ACG CTC GCG CTC TGC GCA GCG CTG AGC CCA GTC ACC
 Met Arg Ile Arg Leu Ala Thr Leu Ala Leu Cys Ala Ala Leu Ser Pro Val Thr

 63 72 81 90 99 108
 TTT GCA GAT AAT GTA ACC GTA CAA ATC GAC GCC GAC GGC GGT AAA AAA CTC ATC
 Phe Ala Asp Asn Val Thr Val Gln Ile Asp Ala Asp Gly Gly Lys Lys Leu Ile

 117 126 135 144 153 162
 AGC CGA GCC CTT TAC GGC ATG AAT AAC TCC AAC GCA GAA AGC CTT ACC GAT ACT
 Ser Arg Ala Leu Tyr Gly Met Asn Asn Ser Asn Ala Glu Ser Leu Thr Asp Thr

 171 180 189 198 207 216
 GAC TGG CAG CGT TTT CGC GAT GCA GGT GTG CGC ATG CTG CGG GAA AAT GGC GGC
 Asp Trp Gln Arg Phe Arg Asp Ala Gly Val Arg Met Leu Arg Glu Asn Gly Gly

 225 234 243 252 261 270
 AAC AAC AGC ACC AAA TAT AAC TGG CAA CTG CAC CTG AGC AGT CAT CCG GAT TGG
 Asn Asn Ser Thr Lys Tyr Asn Trp Gln Leu His Leu Ser Ser His Pro Asp Trp

 279 288 297 306 315 324
 TAC AAC AAT GTC TAC GCC GGC AAC AAC AAC TGG GAC AAC CGG GTA GCC CTG ATT
 Tyr Asn Asn Val Tyr Ala Gly Asn Asn Asn Trp Asp Asn Arg Val Ala Leu Ile

 333 342 351 360 369 378
 CAG GAA AAC CTG CCC GGC GAC ACC ATG TGG GCA TTC CAG CTC ATC CGT AAG
 Gln Glu Asn Leu Pro Gly Ala Asp Thr Met Trp Ala Phe Gln Leu Ile Gly Lys

 387 396 405 414 423 432
 GTC GCG GCG ACT TCT GCC TAC AAC TTT AAC GAT TGG GAA TTC AAC CAG TCG CAA
 Val Ala Ala Thr Ser Ala Tyr Asn Phe Asn Asp Trp Glu Phe Asn Gln Ser Gln

 441 450 459 468 477 486
 TGG TGG ACC GGC GTC GCT CAG AAT CTC GCT GGC GGC CGT GAA CCC AAT CTG GAC
 Trp Trp Thr Gly Val Ala Gln Asn Leu Ala Gly Gly Glu Pro Asn Leu Asp

 495 504 513 522 531 540
 GGC GGC GGC GAA GCG CTG GTT GAA GGA GAC CCC AAT CTC TAC CTC ATG GAT TGG
 Gly Gly Glu Ala Leu Val Glu Gly Asp Pro Asn Leu Tyr Leu Met Asp Trp

 549 558 567 576 585 594
 TCG CCA GCC GAC ACT GTG GGT ATT CTC GAC CAC TGG TTT GGC GTA AAC GGC CTC
 Ser Pro Ala Asp Thr Val Gly Ile Leu Asp His Trp Phe Gly Val Asn Gly Leu

 603 612 621 630 639 648
 GGC GTG CGG CGT GGC AAA GCC AAA TAC TGG AGT ATG GAT AAC GAG CCC GGC ATC
 Gly Val Arg Arg Gly Lys Ala Lys Tyr Trp Ser Met Asp Asn Glu Pro Gly Ile

 657 666 675 684 693 702
 TGG GTT GGC ACC CAC GAC GAT GTA GTG AAA GAA CAA ACC CCG GTA GAA GAT TTC
 Trp Val Gly Thr His Asp Asp Val Val Lys Glu Gln Thr Pro Val Glu Asp Phe

Figure 9

Bankia gouldi endoglucanase (37GP1) (continued)

711	720	729	738	747	756
CTG CAC ACC TAT TTC GAA ACC GCC AAA AAA GCC CGC GCC AAA TTT CCC GGT ATT					
Leu His Thr Tyr Phe Glu Thr Ala Lys Lys Ala Arg Ala Lys Phe Pro Gly Ile					
765	774	783	792	801	810
AAA ATC ACC GGT CCC CTG CCC GCT AAT GAG TGG CAG TGG TAT GCC TGG GCC GGT					
Lys Ile Thr Gly Pro Val Pro Ala Asn Glu Trp Gln Trp Tyr Ala Trp Gly Gly					
819	828	837	846	855	864
TTC TCG GTA CCC CAG GAA CAA GGG TTT ATG AGC TGG ATG GAG TAT TTC ATC AAG					
Phe Ser Val Pro Gln Glu Gln Gly Phe Met Ser Trp Met Glu Tyr Phe Ile Lys					
873	882	891	900	909	918
CGG GTG TCT GAA GAG CAA CGC GCA ACT GGT GTT CCC CTC CTC GAT GTA CTC GAT					
Arg Val Ser Glu Glu Gln Arg Ala Ser Gly Val Arg Leu Leu Asp Val Leu Asp					
927	936	945	954	963	972
CTG CAC TAC TAC CCC GGC GCT TAC AAT GCG GAA GAT ATC GTG CAA TTA CAT CCC					
Leu His Tyr Tyr Pro Gly Ala Tyr Asn Ala Glu Asp Ile Val Gln Leu His Arg					
981	990	999	1008	1017	1026
ACG TTC TTC GAC CGC GAC TTT GTT TCA CTG GAT CCC AAC GGG GTG AAA ATG GTA					
Thr Phe Phe Asp Arg Asp Phe Val Ser Leu Asp Ala Asn Gly Val Lys Met Val					
1035	1044	1053	1062	1071	1080
GAA GGT GGC TGG GAT GAC AGC ATC AAC AAG GAA TAT ATT TTC GGG CGA GTG AAC					
Glu Gly Gly Trp Asp Asp Ser Ile Asn Lys Glu Tyr Ile Phe Gly Arg Val Asn					
1089	1098	1107	1116	1125	1134
GAT TGG CTC GAG GAA TAT ATG GGG CCA GAC CAT GGT GTA ACC CTG GGC TTA ACC					
Asp Trp Leu Glu Tyr Met Gly Pro Asp His Gly Val Thr Leu Gly Leu Thr					
1143	1152	1161	1170	1179	1188
GAA ATG TGC GTG CGC AAT GTG AAT CGG ATG ACT ACC GCC ATC TGG TAT GCC TCC					
Glu Met Cys Val Arg Asn Val Asn Pro Met Thr Thr Ala Ile Trp Tyr Ala Ser					
1197	1206	1215	1224	1233	1242
ATG CTC GCC ACC TTC GCG GAT AAC GGC GTC GAA ATA TTC ACC CCA TGG TGC TCC					
Met Leu Gly Thr Phe Ala Asp Asn Gly Val Glu Ile Phe Thr Pro Trp Cys Trp					
1251	1260	1269	1278	1287	1296
AAC ACC GGA ATG TGG GAA ACA CTC CAC CTC TTC AGC CGC TAC AAC AAA CCT TAT					
Asn Thr Gly Met Trp Glu Thr Leu His Leu Phe Ser Arg Tyr Asn Lys Pro Tyr					
1305	1314	1323	1332	1341	1350
CGG GTC GCC TCC AGC TCC AGT CTT GAA GAG TTT GTC ACC CCC TAC AGC TCC ATT					
Arg Val Ala Ser Ser Ser Leu Glu Glu Phe Val Ser Ala Tyr Ser Ser Ile					
1359	1368	1377	1386	1395	1404
AAC GAA GCA GAA GAC GCC ATG ACG GTA CTT CTG GTG AAT CGT TCC ACT AGC GAC					
Asn Glu Ala Glu Asp Ala Met Thr Val Leu Val Asn Arg Ser Thr Ser Glu					

Figure 9 (Continued)

Bankia gouldi endoglucanase (37GP1) (continued)

1413 1422 1431 1440 1449 1458
ACC CAC ACC GCC ACT GTC GCT ATC GAC GAT TTC CCA CTG GAT GGC CCC TAC CGC
Thr His Thr Ala Thr Val Ala Ile Asp Asp Phe Pro Leu Asp Gly Pro Tyr Arg

1467 1476 1485 1494 1503 1512
ACC CTG CGC TTA CAC AAC CTG CGG GGG GAG GAA ACC TTC GTA TCT CAC CGA GAC
Thr Leu Arg Leu His Asn Leu Pro Gly Glu Thr Phe Val Ser His Arg Asp

1521 1530 1539 1548 1557 1566
AAC GCC CTG GAA AAA GGT ACA GTG CGC GCC AGC GAC AAT ACG GTA ACA CTC GAG
Asn Ala Leu Glu Lys Gly Thr Val Arg Ala Ser Asp Asn Thr Val Thr Leu Glu

1575 1584 1593 1602 1611
TTG CCC CCT CTG TCC GTT ACT GCA ATA TTG CTC AAG GCC CGG CCC TAA 3'
Leu Pro Pro Leu Ser Val Thr Ala Ile Leu Lys Ala Arg Pro ***

Figure 9 (Continued)

Thiopontoya maritima Alpha-D-Glucosidase
Complete Gene Sequence (1-594)

9 18 27 36 45 54
 5 GTG ATC TGT GTG GAA ATA TTC GGA AAC ACC TTC AGA CAG GGA AGA TTC GTT CTC
 Val Ile Cys Val Glu Ile Phe Gly Lys Thr Phe Arg Glu Gly Arg Phe Val Leu

 63 72 81 90 99 108
 AAA GAG AAA AAC TTC ACA GTT GAG TTC GCG GTG GAG AAG ATA CAC CTT GGC TOC
 Lys Glu Lys Asn Phe Thr Val Glu Phe Ala Val Glu Lys Ile His Leu Gly Trp

 117 126 135 144 153 162
 AAG ATC TCC GGC AGG GTG AAG GGA AGT CCG GGA AGG CTT GAG GTC CTT CGA ACC
 Lys Ile Ser Gly Arg Val Lys Gly Ser Pro Gly Arg Leu Glu Val Leu Arg Thr

 171 180 189 198 207 216
 AAA GCA CGG GAA AAG GTC CTT GTG AAC AAC TCG CAG TCC TGG GGA CGG TCC AGG
 Lys Ala Pro Glu Val Leu Val Asn Asn Trp Gln Ser Trp Gly Pro Cys Arg

 225 234 243 252 261 270
 GTG GTC GAT GCC TTT TCT TTC AAA CCA CCT GAA ATA GAT CGG AAC TGG AGA TAC
 Val Val Asp Ala Phe Ser Phe Lys Pro Pro Glu Ile Asp Pro Asn Trp Arg Tyr

 279 288 297 306 315 324
 ACC GCT TCG GTG GTG CCC GAT GTC CTT GAA AGG AAC CTC CAG AGC GAC TAT TTC
 Thr Ala Ser Val Val Pro Asp Val Leu Glu Arg Asn Leu Gln Ser Asp Tyr Phe

 333 342 351 360 369 378
 GTG GCT GAA GAA GGA AAA GTG TAC GGT TTT CTG AGT TCG AAA ATC GCA CAT CCT
 Val Ala Glu Glu Gly Val Tyr Gly Phe Leu Ser Ser Lys Ile Ala His Pro

 387 396 405 414 423 432
 TTC TTC GCT GTG GAA GAT GCG GAA CTT GTG GCA TAC CTC GAA TAT TTC GAT GTC
 Phe Phe Ala Val Glu Asp Gly Glu Leu Val Ala Tyr Leu Glu Tyr Phe Asp Val

 441 450 459 468 477 486
 GAC TTC GAC GAC TTT GTT CCT CTT GAA CCT CTC GTT GTC CTC GAG GAT CCC AAC
 Glu Phe Asp Asp Phe Val Pro Leu Glu Pro Leu Val Val Leu Glu Asp Pro Asn

 496 504 513 522 531 540
 ACA CCC CTT CTT CTG GAG AAA TAC GCG GAA CTC GTC GGA ATG GAA AAC AAC GCG
 Thr Pro Leu Leu Leu Glu Lys Tyr Ala Glu Leu Val Gly Met Glu Asn Asn Ala

 549 558 567 576 585 594
 AGA GTT CCA AAA CAC ACA CCC ACT GGA TCG TCC AGC TCC TAC CAT TAC TTC CTT
 Arg Val Pro Lys His Thr Pro Thr Gly Trp Cys Ser Thr Tyr His Tyr Phe Leu

Figure 10

Thermotoga maritima Alpha-galactosidase
Complete Gene Sequence (2 of 2)

603	612	621	630	639	648
GAT CTC ACT TCG GAA GAG ACC CTC AAG AAC CTC AAG CTC OCG AAC AAT TTC CCG					
Asp Leu Thr Trp Glu Glu Thr Leu Lys Asn Leu Lys Leu Ala Lys Asn Phe Pro					
657	666	675	684	693	702
TTC GAG GTC TTC CAG ATA GAC GAC GCC TAC GAA AAG CAC ATA GGT GAC TGG CTC					
Phe Glu Val Phe Gln Ile Asp Asp Ala Tyr Glu Lys Asp Ile Gly Asp Trp Leu					
711	720	729	738	747	756
GTG ACA AGA GGA GAC TTT CCA TCG GTG GAA GAG ATG GCA AAA GTT ATA GCG GAA					
Val Thr Arg Gly Asp Phe Pro Ser Val Glu Glu Met Ala Lys Val Ile Ala Glu					
765	774	783	792	801	810
AAC GGT TTC ATC CCG GGC ATA TGG ACC GCC CCG TTC AGT GTT TCT GAA ACC TCG					
Asn Gly Phe Ile Pro Gly Ile Trp Thr Ala Pro Phe Ser Val Ser Glu Thr Ser					
819	828	837	846	855	864
GAT GTC TTC AAC GAA CAT CCG GAC TGG GTC AAG GAA AAC CGA GAG CCG AAG					
Asp Val Phe Asn Glu His Pro Asp Trp Val Val Lys Glu Asn Gly Glu Pro Lys					
873	882	891	900	909	918
ATG GCT TAC AGA AAC TCG AAC AAA AAG ATA TAC GCC CTC GAT CTT TCG AAA GAT					
Met Ala Tyr Arg Asn Trp Asn Lys Ile Tyr Ala Leu Asp Leu Ser Lys Asp					
927	936	945	954	963	972
GAG GTT CTG AAC TGG CTT TTC GAT CTC TTC TCA TCT CTG AGA AAG ATG GGC TAC					
Glu Val Leu Asn Trp Leu Phe Asp Ser Ser Leu Arg Lys Met Gly Tyr					
981	990	999	1008	1017	1026
AGG TAC TTC AAG ATC GAC TTT CTC TTC CCC GGT GTC CCA CGA GAA AGA AAA					
Arg Tyr Phe Lys Ile Asp Phe Leu Phe Ala Gly Ala Val Pro Gly Glu Arg Lys					
1035	1044	1053	1062	1071	1080
AAG AAC ATA ACA CCA ATT CAG GCG TTC AGA AAA GGG ATT GAG ACC ATC AGA AAA					
Lys Asn Ile Thr Pro Ile Gln Ala Phe Arg Lys Gly Ile Glu Thr Ile Arg Lys					
1089	1098	1107	1116	1125	1134
GCG GTG GGA GAA GAT TCT TTC ATC CTC CGA TCG GGC TCT CCC CTT CCC GCA					
Ala Val Gly Asp Ser Phe Ile Leu Gly Cys Gly Ser Pro Leu Leu Pro Ala					
1143	1152	1161	1170	1179	1188
GTC CGA TCC GTC GAC GGG ATG AGG ATA GGA CCT GAC ACT CGG CGG TTC TGG GGA					
Val Gly Cys Val Asp Gly Met Arg Ile Gly Pro Asp Thr Ala Pro Phe Trp Gly					

Figure 10 (Continued)

Thiomicrosphaera maritima Alpha-3' nucleosidase
 Complete Gene Sequence (5' to 3')

1197	1206	1215	1224	1233	1242
GAA CAT ATA GAA GAC AAC GCA CCT CCC CCT GCA AGA TOG CGG CTC AGA AAC GCC					
Glu His Ile Glu Asp Asn Gly Ala Pro Ala Ala Arg Trp Ala Leu Arg Asn Ala					
1251	1260	1269	1278	1287	1296
ATA ACG AGG TAC TTC ATG CAC GAC ACG TTC TOG CTG AAC GAC CCC GAC TGT CTG					
Ile Thr Arg Tyr Phe Met His Asp Arg Phe Trp Leu Asn Asp Pro Asp Cys Leu					
1305	1314	1323	1332	1341	1350
ATA CTG AGA GAG GAG AAA ACG GAT CTC ACA CAG AAG GAA AAG GAG CTC TAC TCG					
Ile Leu Arg Glu Glu Lys Thr Asp Leu Thr Gln Lys Glu Lys Glu Leu Tyr Ser					
1359	1368	1377	1386	1395	1404
TAC ACG TGT GGA GTG CTC GAC AAC ATG ATC ATA GAA AGC GAT GAT CTC TCG CTC					
Tyr Thr Cys Gly Val Leu Asp Asn Met Ile Ile Glu Ser Asp Asp Leu Ser Leu					
1413	1422	1431	1440	1449	1458
GTC AGA GAT CAT GGA AAA AAG GTT CTC AAA GAA ACG CTC GAA CTC CTC GGT GGA					
Val Arg Asp His Gly Lys Lys Val Leu Lys Glu Thr Leu Glu Leu Leu Gly Gly					
1467	1476	1485	1494	1503	1512
AGA CCA CGG GTT CAA AAC ATC ATG TCG GAG GAT CTG AGA TAC GAG ATC GTC TCG					
Arg Pro Arg Val Gln Asn Ile Met Ser Glu Asp Leu Arg Tyr Glu Ile Val Ser					
1521	1530	1539	1548	1557	1566
TCT GGC ACT CTC TCA GCA AAC GTC AAG ATC GTG GTC GAT CTG AAC AGC AGA GAG					
Ser Gly Thr Leu Ser Gly Asn Val Lys Ile Val Val Asp Leu Asn Ser Arg Glu					
1575	1584	1593	1602	1611	1620
TAC CAC CTG GAA AAA GAA GGA AAG TCT TCC CTG AAA AAA AGA GTC GTC AAA AGA					
Tyr His Leu Glu Lys Glu Gly Lys Ser Ser Leu Lys Lys Arg Val Val Lys Arg					
1629	1638	1647	1656	1665	
GAA GAC CGA AGA AAC TTC TAC TTC TAC GAA GAC CCT GAG AGA GAA TGA 3'					
Glu Asp Gly Arg Asn Phe Tyr Phe Tyr Glu Glu Gly Glu Arg Glu ***					

Figure 10 (Continued)

Thermotoga maritima β -mannanase (6GP2)

9	18	27	36	45	54												
ATG	GGG	ATT	GGT	GGC	GAC	GAC	TCC	TGG	AGC	CCG	TCA	GTA	TCG	GGG	GAA	TTC	CTT
Met	Gly	Ile	Gly	Gly	Asp	Asp	Ser	Trp	Ser	Pro	Ser	Val	Ser	Ala	Glu	Phe	Leu
63	72	81	90	99	108												
TTA	TTC	ATC	GTT	GAG	CTC	TCT	TTC	GTT	CTC	TTT	GCA	AGT	GAC	GAG	TTC	GTG	AAA
Leu	Leu	Ile	Val	Glu	Leu	Ser	Phe	Val	Leu	Phe	Ala	Ser	Asp	Glu	Phe	Val	Lys
117	126	135	144	153	162												
CTG	GAA	AAC	GGA	AAA	TTC	GCT	CTG	AAC	GGA	AAA	GAA	TTC	AGA	TTC	ATT	GGA	AGC
Val	Glu	Asn	Gly	Lys	Phe	Ala	Leu	Asn	Gly	Lys	Glu	Phe	Arg	Phe	Ile	Gly	Ser
171	180	189	198	207	216												
AAC	AAC	TAC	TAC	ATG	CAC	TAC	AAG	AGC	AAC	GGA	ATG	ATA	GAC	AGT	GTT	CTG	GAG
Asn	Asn	Tyr	Tyr	Met	His	Tyr	Lys	Ser	Asn	Gly	Met	Ile	Asp	Ser	Val	Leu	Glu
225	234	243	252	261	270												
AGT	GCC	AGA	GAC	ATG	GGT	ATA	AAG	GTC	CTC	AGA	ATC	TGG	GGT	TTC	CTC	GAC	GGG
Ser	Ala	Arg	Asp	Met	Gly	Ile	Lys	Val	Leu	Arg	Ile	Trp	Gly	Phe	Leu	Asp	Gly
279	288	297	306	315	324												
GAG	AGT	TAC	TGC	AGA	GAC	AAG	AAC	ACC	TAC	ATG	CAT	CCT	GAG	CCC	GGT	GTT	TTC
Glu	Ser	Tyr	Cys	Arg	Asp	Lys	Asn	Thr	Tyr	Met	His	Pro	Glu	Pro	Gly	Val	Phe
333	342	351	360	369	378												
GGG	GTC	CCA	GAA	GGG	ATA	TCG	AAC	GCC	CAG	AGC	GGT	TTC	GAA	AGA	CTC	GAC	TAC
Gly	Val	Pro	Glu	Gly	Ile	Ser	Asn	Ala	Gln	Ser	Gly	Phe	Glu	Arg	Leu	Asp	Tyr
387	396	405	414	423	432												
ACA	GTT	GCG	AAA	GCG	AAA	GAA	CTC	GGT	ATA	AAA	CTT	GTC	ATT	GTT	CTT	GTG	AAC
Thr	Val	Ala	Lys	Ala	Lys	Glu	Leu	Gly	Ile	Lys	Leu	Val	Ile	Val	Leu	Val	Asn
441	450	459	468	477	486												
AAC	TGG	GAC	GAC	TTC	GGT	GGA	ATG	AAC	CAG	TAC	GTG	AGG	TGG	TTT	GGA	GGA	ACC
Asn	Trp	Asp	Asp	Phe	Gly	Met	Aan	Gln	Tyr	Val	Arg	Trp	Phe	Gly	Gly	Thr	
495	504	513	522	531	540												
CAT	CAC	GAC	GAT	TTC	TAC	AGA	GAT	GAG	AAG	ATC	AAA	GAA	GAG	TAC	AAA	AAG	TAC
His	His	Asp	Asp	Phe	Tyr	Arg	Asp	Glu	Lys	Ile	Lys	Glu	Glu	Tyr	Lys	Lys	Tyr

Figure 11

Thermotoga maritima β -mannanase (continued) (6 Gf2)

549	558	567	576	585	594
GTC TCC TTT CTC GTA AAC CAT GTC AAT ACC TAC ACG GGA GTT CCT TAC AGG GAA					
---	---	---	---	---	---
Val Ser Phe Leu Val Asn His Val Asn Thr Tyr Thr Gly Val Pro Tyr Arg Glu					
603	612	621	630	639	648
GAG CCC ACC ATC ATG GCC TGG GAG CTT GCA AAC GAA CCG CCC TGT GAG ACG GAC					
---	---	---	---	---	---
Glu Pro Thr Ile Met Ala Trp Glu Leu Ala Asn Glu Pro Arg Cys Glu Thr Asp					
657	666	675	684	693	702
AAA TCG GGG AAC ACG CTC GTT GAG TGG GTG AAG GAG ATG AGC TCC TAC ATA AAG					
---	---	---	---	---	---
Lys Ser Gly Asn Thr Leu Val Glu Trp Val Lys Glu Met Ser Ser Tyr Ile Lys					
711	720	729	738	747	756
AGT CTG GAT CCC AAC CAC CTC GTG GCT GTG GGG GAC GAA GGA TTC TTC AGC AAC					
---	---	---	---	---	---
Ser Leu Asp Pro Asn His Leu Val Ala Val Gly Asp Glu Gly Phe Phe Ser Asn					
765	774	783	792	801	810
TAC GAA CGA TTC AAA CCT TAC GGT GGA GAA GCC GAG TGG GCC TAC AAC GGC TGG					
---	---	---	---	---	---
Tyr Glu Gly Phe Lys Pro Tyr Gly Gly Glu Ala Glu Trp Ala Tyr Asn Gly Trp					
819	828	837	846	855	864
TCC GGT GTT GAC TGG AAG AAG CTC CTT TCG ATA GAG ACG GTG GAC TTC GGC ACG					
---	---	---	---	---	---
Ser Gly Val Asp Trp Lys Lys Leu Leu Ser Ile Glu Thr Val Asp Phe Gly Thr					
873	882	891	900	909	918
TTC CAC CTC TAT CCG TCC CAC TGG GGT GTC AGT CCA GAG AAC TAT GCC CAG TGG					
---	---	---	---	---	---
Phe His Leu Tyr Pro Ser His Trp Gly Val Ser Pro Glu Asn Tyr Ala Gln Trp					
927	936	945	954	963	972
GGA GCG AAG TGG ATA GAA GAC CAC ATA AAG ATC GCA AAA GAG ATC GGA AAA CCC					
---	---	---	---	---	---
Gly Ala Lys Trp Ile Glu Asp His Ile Lys Ile Ala Lys Glu Ile Gly Lys Pro					
981	990	999	1008	1017	1026
GTT GTT CTG GAA GAA TAT GGA ATT CCA AAG AGT GCG CCA GTT AAC AGA ACG GCC					
---	---	---	---	---	---
Val Val Leu Glu Glu Tyr Gly Ile Pro Lys Ser Ala Pro Val Asn Arg Thr Ala					
1035	1044	1053	1062	1071	1080
ATC TAC AGA CTC TGG AAC GAT CTG GTC TAC GAT CTC GGT GGA GAT GGA GCG ATG					
---	---	---	---	---	---
Ile Tyr Arg Leu Trp Asn Asp Leu Val Tyr Asp Leu Gly Gly Asp Gly Ala Met					

Figure 11 (Continued)

Thermotoga maritima β -mannanase (6GP2) (continued)

1089	1098	1107	1116	1125	1134
TTC	TGG	ATG	CTC	GCG	GGA
ATC	GGG	GAA	GGT	TCG	GAC
GAC	AGA	GAC	GAG	AGA	GCG
GAC	TAC	GAC	GAC	GAC	TAC
---	---	---	---	---	---
Phe	Trp	Met	Leu	Ala	Gly
Ile	Gly	Gly	Gly	Ser	Asp
Asp	Arg	Asp	Glu	Arg	Gly
Arg	Asp	Glu	Tyr		
1143	1152	1161	1170	1179	1188
TAT	CCG	GAC	TAC	GAC	GGT
TTC	AGA	ATA	GTG	AAC	GAC
GAC	AGA	GAA	GAC	GAC	AGT
---	---	---	---	---	---
Tyr	Pro	Asp	Tyr	Asp	Gly
Asp	Tyr	Asp	Gly	Phe	Arg
Arg	Ile	Val	Asn	Asp	Asp
Asp			Ser	Pro	Glu
1197	1206	1215	1224	1233	1242
CTG	ATA	AGA	GAA	TAC	GCG
AAG	TAC	GGC	AAG	CTG	TTC
ACA	AAC	ACA	ACA	GGT	GAA
GAC	GAC	GAC	GAC	GAA	GAC
---	---	---	---	---	---
Leu	Ile	Arg	Glu	Tyr	Ala
Lys	Leu	Phe	Asn	Thr	Gly
Asp				Glu	Asp
Asp				Ile	Arg
Arg				Glu	Asp
1251	1260	1269	1278	1287	1296
ACC	TGC	TCT	TTC	ATC	CTT
ATC	GGT	ATA	GGC	GAG	ATC
CTT	TTC	AAA	GAC	AAA	AAA
CCA	AGC	GCA	GCC	AAG	AAG
---	---	---	---	---	---
Thr	Cys	Ser	Phe	Ile	Lys
Asp	Leu	Pro	Lys	Asp	Gly
Val				Gly	Asp
				Met	Ile
				Lys	Lys
				Thr	Val
				Glu	Glu
1305	1314	1323	1332	1341	1350
GTG	AGG	GCT	GGT	GTT	TTC
GCT	TTC	TTC	GAC	TAC	ATC
TTC	GGT	AGC	TAC	AGC	ATA
---	---	---	---	---	---
Val	Arg	Ala	Gly	Val	Asp
Asp	Tyr	Ser	Asn	Thr	Phe
Tyr	Ser	Asn	Thr	Glu	Lys
1359	1368	1377	1386	1395	1404
GTC	GAA	GAT	CTG	GTT	TTT
GAA	GAT	CTG	GTT	TTC	GAA
AAT	TTC	TTT	GAA	ATA	ATA
GAG	GAC	GAC	ATA	GAG	GTC
---	---	---	---	---	---
Val	Glu	Asp	Leu	Val	Phe
Asp	Tyr	Glu	Asn	Glu	Ile
Tyr	Ser	Asn	Thr	Glu	Gly
Ser	Asn	Thr	Phe	Ile	Tyr
1413	1422	1431	1440	1449	1458
GTC	TTT	GAT	CTC	GAC	ACA
GAT	GAT	CTC	GAC	ACC	ACC
CTC	GAC	ACA	ACC	CGG	ATC
GAC	ATC	ATA	CGG	GAT	TTC
---	---	---	---	---	---
Gly	Phe	Asp	Leu	Asp	Thr
Asp	Asp	Thr	Thr	Arg	Ile
Leu	Asp	Asp	Asp	Pro	Pro
				Gly	Glu
				Glu	His
				His	Ile
				Glu	Gly
1467	1476	1485	1494	1503	1512
GAA	GGC	CAC	TTT	CAG	GGG
GGC	CAC	TTT	CAG	GGG	AAA
CAC	AAA	ACG	AAA	ACG	GTG
TTT	ACG	GTG	AAA	TCT	TTC
---	---	---	---	---	---
Glu	Gly	His	Phe	Gln	Gly
Gly	Lys	Asp	Asp	Lys	Lys
His	Thr	Asp	Ser	Ile	Ala
Phe	Val	Ser	Ile	Lys	Lys
Gln	Lys	Ser	Lys	Ala	Val
1521	1530	1539	1548	1557	1566
AAC	GAA	GCA	CGG	TAC	GTG
GAA	GCA	CGG	TAC	GTG	CTC
TAC	GTG	CTC	GCA	GAG	GAG
---	---	---	---	---	---
Asn	Glu	Ala	Arg	Tyr	Val
Glu	Ala	Ala	Tyr	Val	Leu
Asp	Asp	Asp	Asp	Asp	Glu
Asp	Asp	Asp	Asp	Asp	Glu
1575	1584	1593	1602	1611	1620
GTG	AAA	AAC	TGG	TGG	AAC
AAA	AAC	TGG	TGG	AAC	AGC
AAC	TGG	AAC	AGC	AGC	GGA
TGG	AAC	AGC	AGC	ACC	ACC
AAC	AGC	GGA	GGA	TGG	CAG
AGC	GGA	AGA	AGA	CAG	GCA
GGA	AGA	GAA	GAA	GAG	GAG
AGA	GAA	GAA	GAA	TTC	TTC
GAG	GAA	GAA	GAG	GGG	GGG
GAG	GAA	GAA	GAG	TCA	TCA
GAG	GAA	GAA	GAG	CCT	CCT
GAG	GAA	GAA	GAG	GAC	GAC
---	---	---	---	---	---
Val	Lys	Asn	Trp	Trp	Asn
				Ser	Ser
				Gly	Gly
				Thr	Thr
				Trp	Gln
				Asp	Asp

Figure 11 (Continued)

Thermotoga maritima β -mannanase (6G12) (continued) (6G12)

1629	1638	1647	1656	1665	1674
ATT	GAA	TGG	AAC	GGT	GAG
---	---	---	---	---	---
Ile	Glu	Trp	Asn	Gly	Glu
1683	1692	1701	1710	1719	1728
CCC	CGA	AAG	ACC	GAC	TGG
---	---	---	---	---	---
Pro	Gly	Lys	Ser	Asp	Trp
1737	1746	1755	1764	1773	1782
TCA	GAA	TGT	GAG	ATC	CTC
---	---	---	---	---	---
Ser	Glu	Cys	Glu	Ile	Leu
1791	1800	1809	1818	1827	1836
AAG	GGA	AGG	TTG	AGG	CCG
---	---	---	---	---	---
Lys	Gly	Arg	Leu	Arg	Pro
1845	1854	1863	1872	1881	1890
CTC	GAC	ATG	AAC	AAC	GCG
---	---	---	---	---	---
Leu	Asp	Met	Asn	Asn	Ala
1899	1908	1917	1926	1935	1944
AAA	GAG	TAC	AGA	AGA	TTC
---	---	---	---	---	---
Lys	Glu	Tyr	Arg	Arg	Phe
1953	1962	1971	1980	1989	1998
AAA	GAA	CTT	CAC	ATA	GGG
---	---	---	---	---	---
Lys	Glu	Leu	His	Ile	Gly
2007	2016	2025	2034	2043	
TTC	ATC	GAT	AAT	GTG	AGA
---	---	---	---	---	---
Phe	Ile	Asp	Asn	Val	Arg

Figure 11 (Continued)

AEPII 1a β -mannosidase (63GB1)

9	18	27	36	45	54
ATG CTA CCA GAA GAG TTC CTA TGG GGC GTT GGG CAG TCA GGC TTT CAG TTC GAA					
Met Leu Pro Glu Glu Phe Leu Trp Gly Val Gly Gln Ser Gly Phe Gln Phe Glu					
63	72	81	90	99	108
ATG GGC GAC AAG CTC AGG AGG CAC ATC GAT CCA AAT ACC GAC TGG TGG AAG TGG					
Met Gly Asp Lys Leu Arg Arg His Ile Asp Pro Asn Thr Asp Trp Trp Lys Trp					
117	126	135	144	153	162
GTT CGC GAT CCT TTC AAC ATA AAA AAG GAG CTT GTG AGT GGG GAC CTT CCC GAG					
Val Arg Asp Pro Phe Asn Ile Lys Lys Glu Leu Val Ser Gly Asp Leu Pro Glu					
171	180	189	198	207	216
GAC GGC ATC AAC AAC TAC GAA CTT TTT GAA AAC GAT CAC AAG CTC GCT AAA GGC					
Asp Gly Ile Asn Asn Tyr Glu Leu Phe Glu Asn Asp His Lys Leu Ala Lys Gly					
225	234	243	252	261	270
CTT GGA CTC AAC GCA TAC AGG ATT GGA ATA GAG TGG AGC AGA ATC TTT CCC TGG					
Leu Gly Leu Asn Ala Tyr Arg Ile Gly Ile Glu Trp Ser Arg Ile Phe Pro Trp					
279	288	297	306	315	324
CCG ACG TGG ACG GTC GAT ACC GAG GTC GAG TTC GAC ACT TAC GGT TTA GTA AAG					
Pro Thr Trp Thr Val Asp Thr Glu Val Glu Phe Asp Thr Tyr Gly Leu Val Lys					
333	342	351	360	369	378
GAC GTT AAG ATA GAC AAG TCC ACC CTT GCT GAA CTC GAC AGG CTG GCC AAC AAG					
Asp Val Lys Ile Asp Lys Ser Thr Leu Ala Glu Leu Asp Arg Leu Ala Asn Lys					
387	396	405	414	423	432
GAG GAG GTA ATG TAC TAC AGG CGC GTT ATT CAG CAT TTG AGG GAG CTC GGC TTC					
Glu Glu Val Met Tyr Tyr Arg Arg Val Ile Gln His Leu Arg Glu Leu Gly Phe					
441	450	459	468	477	486
AAG GTC TTC GTT AAC CTC AAC CAC TTC ACG CTT CCA ATA TGG CTC CAC GAC CCG					
Lys Val Phe Val Asn Leu Asn His Phe Thr Leu Pro Ile Trp Leu His Asp Pro					
495	504	513	522	531	540
ATA GTG GCA AGG GAG AAG GCC CTC ACA AAC GAC AGA ATC GGC TGG GTC TCC CAG					
Ile Val Ala Arg Glu Lys Ala Leu Thr Asn Asp Arg Ile Gly Trp Val Ser Gln					

Figure 12

AEPPI 1a β -mannosidase (630B1) (continued)

549	558	567	576	585	594
ACG ACA GTT CTT GAG TTT GCC AAG TAT CCT GCT TAC ATC GCC CAT GCG CTC GGA					
--- --- --- --- --- ---					
Arg Thr Val Val Glu Phe Ala Lys Tyr Ala Ala Tyr Ile Ala His Ala Leu Gly					
603	612	621	630	639	648
GAC CTC GTG GAC ACA TCG AGC ACC TTC AAC GAA CCT ATG GTA GTT GTG GAG CTC					
--- --- --- --- --- ---					
Asp Leu Val Asp Thr Trp Ser Thr Phe Asn Glu Pro Met Val Val Val Glu Leu					
657	666	675	684	693	702
GGC TAC CTC GCC CCC TAC TCA GGA TTT CCC CCG GGA GTC ATG AAC CCC GAG GCC					
--- --- --- --- --- ---					
Gly Tyr Leu Ala Pro Tyr Ser Gly Phe Pro Pro Gly Val Met Asn Pro Glu Ala					
711	720	729	738	747	756
GCG AAG CTG GCG ATC CTC AAC ATG ATA AAC GCG CAC GCC TTG GCA TAT AAG ATG					
--- --- --- --- --- ---					
Ala Lys Leu Ala Ile Leu Asn Met Ile Asn Ala His Ala Leu Ala Tyr Lys Met					
765	774	783	792	801	810
ATA AAG AGG TTC GAC ACC AAG AAG GCC GAT GAG GAT AGC AAG TCC CCT GCG GAC					
--- --- --- --- --- ---					
Ile Lys Arg Phe Asp Thr Lys Lys Ala Asp Glu Asp Ser Lys Ser Pro Ala Asp					
819	828	837	846	855	864
GTT GGC ATA ATT TAC AAC AAC ATC GGT GTT GCC TAC CCT AAA GAC CCT AAC GAT					
--- --- --- --- --- ---					
Val Gly Ile Ile Tyr Asn Asn Ile Gly Val Ala Tyr Pro Lys Asp Pro Asn Asp					
873	882	891	900	909	918
CCC AAG GAC GTT AAA GCA GCC GAA AAC GAC AAC TAC TTC CAC ACC GGA CTG TTC					
--- --- --- --- --- ---					
Pro Lys Asp Val Lys Ala Ala Glu Asn Asp Asn Tyr Phe His Ser Gly Leu Phe					
927	936	945	954	963	972
TTT GAT GCC ATC CAC AAG GGT AAG CTC AAC ATA GAG TTC GAC GGC GAA AAC TTT					
--- --- --- --- --- ---					
Phe Asp Ala Ile His Lys Gly Lys Leu Asn Ile Glu Phe Asp Gly Glu Asn Phe					
981	990	999	1008	1017	1026
GTA AAA GTT AGA CAC CTA AAA GCC AAT GAC TGG ATA GGC CTC AAC TAC TAC ACC					
--- --- --- --- --- ---					
Val Lys Val Arg His Leu Lys Gly Asn Asp Trp Ile Gly Leu Asn Tyr Tyr Thr					
1035	1044	1053	1062	1071	1080
CGC GAG GTT GTT AGA TAT TCG GAG CCC AAG TTC CCA AGT ATA CCC CTC ATA TCC					
--- --- --- --- --- ---					
Arg Glu Val Val Arg Tyr Ser Glu Pro Lys Phe Pro Ser Ile Pro Leu Ile Ser					

Figure 12 (Continued)

AEPII 1a β -mannosidase (63GB1) (continued)

1089	1098	1107	1116	1125	1134												
TTC	AAG	GGC	GTT	CCC	AAC	TAC	GGC	TAC	TCC	TGC	AGG	CCC	GGC	ACG	ACC	TCC	GCC
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Phe	Lys	Gly	Val	Pro	Asn	Tyr	Gly	Tyr	Ser	Cys	Arg	Pro	Gly	Thr	Thr	Ser	Ala
1143	1152	1161	1170	1179	1188												
GAT	GGC	ATG	CCC	GTC	AGC	GAT	ATC	GGC	TGG	GAA	GTC	TAT	CCC	CAG	GGA	ATC	TAC
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Asp	Gly	Met	Pro	Val	Ser	Asp	Ile	Gly	Trp	Glu	Val	Tyr	Pro	Gln	Gly	Ile	Tyr
1197	1206	1215	1224	1233	1242												
GAC	TCG	ATA	GTC	GAG	GCC	ACC	AAG	TAC	AGT	GTT	CCT	GTT	TAC	GTC	ACC	GAG	AAC
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Asp	Ser	Ile	Val	Glu	Ala	Thr	Lys	Tyr	Ser	Val	Pro	Val	Tyr	Val	Thr	Glu	Asn
1251	1260	1269	1278	1287	1296												
GCT	GTT	GCG	GAT	TCC	GGG	GAC	ACG	CTG	AGG	CCA	TAC	TAC	ATA	GTC	AGC	CAC	GTC
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Gly	Val	Ala	Asp	Ser	Ala	Asp	Thr	Leu	Arg	Pro	Tyr	Tyr	Ile	Val	Ser	His	Val
1305	1314	1323	1332	1341	1350												
TCA	AAG	ATA	GAG	GAA	GCC	ATT	GAG	AAT	GGA	TAC	CCC	GTA	AAA	GGC	TAC	ATG	TAC
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Ser	Lys	Ile	Glu	Glu	Ala	Ile	Glu	Asn	Gly	Tyr	Pro	Val	Lys	Gly	Tyr	Met	Tyr
1359	1368	1377	1386	1395	1404												
TGG	GCG	CTT	ACG	GAT	AAC	TAC	GAG	TGG	GCC	CTC	GGC	TTC	AGC	ATG	AGG	TTT	GGT
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Trp	Ala	Leu	Thr	Asp	Asn	Tyr	Glu	Trp	Ala	Leu	Gly	Phe	Ser	Met	Arg	Phe	Gly
1413	1422	1431	1440	1449	1458												
CTC	TAC	AAG	GTC	GAC	CTC	ATC	TCC	AAG	GAG	AGG	ATC	CCG	AGG	GAG	AGA	AGC	GTT
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	
Leu	Tyr	Lys	Val	Asp	Leu	Ile	Ser	Lys	Glu	Arg	Ile	Pro	Arg	Glu	Arg	Ser	Val
1467	1476	1485	1494	1503	1512												
GAG	ATA	TAT	CGC	AGG	ATA	GTG	CAG	TCC	AAC	GGT	GTT	CCT	AAG	GAT	ATC	AAA	GAG
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Glu	Ile	Tyr	Arg	Arg	Ile	Val	Gln	Ser	Asn	Gly	Val	Pro	Lys	Asp	Ile	Lys	Glu
1521	1530	1539															
GAG	TTC	CTG	AAG	GGT	GAG	GAG	AAA	TGA	3'								
---	---	---	---	---	---	---	---	---	---								
Glu	Phe	Leu	Lys	Gly	Glu	Glu	Lys	***									

Figure 12 (Continued)

OC1/4V Endoglucanase (33GP1)

9 18 27 36 45 54
 ATG GTA GAA AGA CAC TTC AGA TAT GTT CTT ATT TGC ACC CTG TTT CTT GTT ATG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Met Val Glu Arg His Phe Arg Tyr Val Leu Ile Cys Thr Leu Phe Leu Val Met

 63 72 81 90 99 108
 CTC CTA ATC TCA TCC ACT CAG TGT GGA AAA AAT GAA CCA AAC AAA AGA GTG AAT
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Leu Leu Ile Ser Ser Thr Gln Cys Gly Lys Asn Glu Pro Asn Lys Arg Val Asn

 117 126 135 144 153 162
 AGC ATG GAA CAG TCA GTT GCT GAA AGT GAT AGC AAC TCA GCA TTT GAA TAC AAC
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Ser Met Glu Gln Ser Val Ala Glu Ser Asp Ser Asn Ser Ala Phe Glu Tyr Asn

 171 180 189 198 207 216
 AAA ATG GTA GGT AAA GGA GTA AAT ATT GGA AAT GCT TTA GAA GCT CCT TTC GAA
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Lys Met Val Gly Lys Val Asn Ile Gly Asn Ala Leu Glu Ala Pro Phe Glu

 225 234 243 252 261 270
 GGA GCT TGG GGA GTA AGA ATT GAG GAT GAA TAT TTT GAG ATA ATA AAG AAA AGG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Gly Ala Trp Gly Val Arg Ile Glu Asp Glu Tyr Phe Glu Ile Ile Lys Lys Arg

 279 288 297 306 315 324
 GGA TTT GAT TCT GTT AGG ATT CCC ATA AGA TGG TCA GCA CAT ATA TCC GAA AAG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Gly Phe Asp Ser Val Arg Ile Pro Ile Arg Trp Ser Ala His Ile Ser Glu Lys

 333 342 351 360 369 378
 CCA CCA TAT GAT ATT GAC AGG AAT TTC CTC GAA AGA GTT AAC CAT GTT GTC GAT
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Pro Pro Tyr Asp Ile Asp Arg Asn Phe Leu Glu Arg Val Asn His Val Val Asp

 387 396 405 414 423 432
 AGG GCT CTT GAG AAT AAT TTA ACA GTA ATC ATC AAT ACG CAC CAT TTT GAA GAA
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Arg Ala Leu Glu Asn Asn Leu Thr Val Ile Ile Asn Thr His His Phe Glu Glu

 441 450 459 468 477 486
 CTC TAT CAA GAA CCG GAT AAA TAC GGC GAT GTT TTG GTG GAA ATT TGG AGA CAG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Leu Tyr Gln Glu Pro Asp Lys Tyr Gly Asp Val Leu Val Glu Ile Trp Arg Gln

 495 504 513 522 531 540
 ATT GCA AAA TTC TTT AAA GAT TAC CCG GAA AAT CTG TTC TTT GAA ATC TAC AAC
 --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Ile Ala Lys Phe Phe Lys Asp Tyr Pro Glu Asn Leu Phe Phe Glu Ile Tyr Asn

Figure 13

OC1/6V Endoglucanase (33GP1) (continued)

549 558 567 576 585 594
 GAG CCT CAG AAC TTG ACA GCT GAA AAA TGG AAC GCA CTT TAT CCA AAA GTC
 Glu Pro Ala Gln Asn Leu Thr Ala Glu Lys Trp Asn Ala Leu Tyr Pro Lys Val
 603 612 621 630 639 648
 CTC AAA GTT ATC AGG GAG AGC AAT CCA ACC CGG ATT GTC ATT ATC GAT GCT CCA
 Leu Lys Val Ile Arg Glu Ser Asn Pro Thr Arg Ile Val Ile Ile Asp Ala Pro
 657 666 675 684 693 702
 AAC TGG GCA CAC TAT AGC GCA GTG AGA AGT CTA AAA TTA GTC AAC GAC AAA CGC
 Asn Trp Ala His Tyr Ser Ala Val Arg Ser Leu Lys Leu Val Asn Asp Lys Arg
 711 720 729 738 747 756
 ATC ATT GTT TCC TTC CAT TAC TAC GAA CCT TTC AAA TTC ACA CAT CAG CGT GCC
 Ile Ile Val Ser Phe His Tyr Tyr Glu Pro Phe Lys Phe Thr His Gln Gly Ala
 765 774 783 792 801 810
 GAA TGG GTT AAT CCC ATC CCA CCT GTT AGG GTT AAG TGG AAT GGC GAG GAA TGG
 Glu Trp Val Asn Pro Ile Pro Pro Val Arg Val Lys Trp Asn Gly Glu Trp
 819 828 837 846 855 864
 GAA ATT AAC CAA ATC AGA AGT CAT TTC AAA TAC GTG AGT GAC TGG GCA AAG CAA
 Glu Ile Asn Gln Ile Arg Ser His Phe Lys Tyr Val Ser Asp Trp Ala Lys Gln
 873 882 891 900 909 918
 AAT AAC GTA CCA ATC TTT CTT GGT GAA TTC GGT GCT TAT TCA AAA GCA GAC ATG
 Asn Asn Val Pro Ile Phe Leu Gly Glu Phe Gly Ala Tyr Ser Lys Ala Asp Met
 927 936 945 954 963 972
 GAC TCA AGG GTT AAG TGG ACC GAA AGT GTG AGA AAA ATG GCG GAA GAA TTT GGA
 Asp Ser Arg Val Lys Trp Thr Glu Ser Val Arg Lys Met Ala Glu Glu Phe Gly
 981 990 999 1008 1017 1026
 TTT TCA TAC GCG TAT TGG GAA TTT TGT GCA GGA TTT GGC ATA TAC GAT AGA TGG
 Phe Ser Tyr Ala Tyr Trp Glu Phe Cys Ala Gly Phe Gly Ile Tyr Asp Arg Trp
 1035 1044 1053 1062 1071 1080
 TCT CAA AAC TGG ATC GAA CCA TTG GCA ACA GCT GTG GTT GGC ACA GGC AAA GAG
 Ser Gln Asn Trp Ile Glu Pro Leu Ala Thr Ala Val Val Gly Thr Gly Lys Glu

TAA 3

Figure 13 (Continued)

Thermotoga maritima pullulanase (6GP3)

9 18 27 36 45 54
 5 ATG GAT CTT ACA AAG GTG GGG ATC ATA GTG AGG CTG AAC GAG TGG CAG GCA AAA
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Met Asp Leu Thr Lys Val Gly Ile Ile Val Arg Leu Asn Glu Trp Gln Ala Lys

 63 72 81 90 99 108
 GAC GTG GCA AAA GAC AGG TTC ATA GAG ATA AAA GAC GGA AAG GCT GAA GTG TGG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Asp Val Ala Lys Asp Arg Phe Ile Glu Ile Lys Asp Gly Lys Ala Glu Val Trp

 117 126 135 144 153 162
 ATA CTC CAG GGA GTG GAA GAG ATT TTC TAC GAA AAA CCA GAC ACA TCT CCC AGA
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Ile Leu Gln Gly Val Glu Glu Ile Phe Tyr Glu Lys Pro Asp Thr Ser Pro Arg

 171 180 189 198 207 216
 ATC TTC TTC GCA CAG GCA AGG TCG AAC AAG GTG ATC GAG GCT TTT CTG ACC AAT
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Ile Phe Phe Ala Gln Ala Arg Ser Asn Lys Val Ile Glu Ala Phe Leu Thr Asn

 225 234 243 252 261 270
 CCT GTG GAT ACG AAA AAG AAA GAA CTC TTC AAG GTT ACT GTT GAC GGA AAA GAG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Pro Val Asp Thr Lys Lys Glu Leu Phe Lys Val Thr Val Asp Gly Lys Glu

 279 288 297 306 315 324
 ATT CCC GTC TCA AGA GTG GAA AAG GCC GAT CCC ACG GAC ATA GAC GTG ACG AAC
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Ile Pro Val Ser Arg Val Glu Lys Ala Asp Pro Thr Asp Ile Asp Val Thr Asn

 333 342 351 360 369 378
 TAC GTG AGA ATC GTC CTT TCT GAA TCC CTG AAA GAA GAA GAC CTC AGA AAA GAC
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Tyr Val Arg Ile Val Leu Ser Glu Ser Leu Lys Glu Glu Asp Leu Arg Lys Asp

 387 396 405 414 423 432
 GTG GAA CTG ATC ATA GAA GGT TAC AAA CCG GCA AGA GTC ATC ATG ATG GAG ATC
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Val Glu Leu Ile Ile Glu Gly Tyr Lys Pro Ala Arg Val Ile Met Met Glu Ile

 441 450 459 468 477 486
 CTG GAC GAC TAC TAT TAC GAT GGA GAG CTC GGA GCC GTA TAT TCT CCA GAG AAG
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Leu Asp Asp Tyr Tyr Asp Gly Glu Leu Gly Ala Val Tyr Ser Pro Glu Lys

 495 504 513 522 531 540
 ACG ATA TTC AGA GTC TGG TCC CCC GTP TCT AAG TGG GTA AAG GTG CTT CTC TTC
 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Thr Ile Phe Arg Val Trp Ser Pro Val Ser Lys Trp Val Lys Val Leu Leu Phe

Figure 14

Thermotoga maritima Pullulanase (6GP3) (continued)

549	558	567	576	585	594
AAA AAC GGA GAA GAC ACA GAA CCG TAC CAG GTT GTG AAC ATG GAA TAC AAG GGA					
---	---	---	---	---	---
Lys Asn Gly Glu Asp Thr Glu Pro Tyr Gln Val Val Asn Met Glu Tyr Lys Gly					
603	612	621	630	639	648
AAC GGG GTC TGG GAA GCG GTT GTT GAA GGC GAT CTC GAC GGA GTG TTC TAC CTC					
---	---	---	---	---	---
Asn Gly Val Trp Glu Ala Val Val Glu Gly Asp Leu Asp Gly Val Phe Tyr Leu					
657	666	675	684	693	702
TAT CAG CTG GAA AAC TAC CGA AAG ATC AGA ACA ACC GTC GAT CCT TAT TCG AAA					
---	---	---	---	---	---
Tyr Gln Leu Glu Asn Tyr Gly Lys Ile Arg Thr Thr Val Asp Pro Tyr Ser Lys					
711	720	729	738	747	756
GCG GTT TAC GCA AAC AAC CAA GAG AGC CCC GTT GTG AAT CTT GCC AGG ACA AAC					
---	---	---	---	---	---
Ala Val Tyr Ala Asn Asn Gln Glu Ser Ala Val Val Asn Leu Ala Arg Thr Asn					
765	774	783	792	801	810
CCA GAA GGA TGG GAA AAC GAC AGG GGA CCG AAA ATC GAA GGA TAC GAA GAC GCG					
---	---	---	---	---	---
Pro Glu Gly Trp Glu Asn Asp Arg Gly Pro Lys Ile Glu Gly Tyr Glu Asp Ala					
819	828	837	846	855	864
ATA ATC TAT GAA ATA CAC ATA GCG GAC ATC ACA GGA CTC GAA AAC TCC GGG GTA					
---	---	---	---	---	---
Ile Ile Tyr Glu Ile His Ile Ala Asp Ile Thr Gly Leu Glu Asn Ser Gly Val					
873	882	891	900	909	918
AAA AAC AAA GGC CTC TAT CTC GGG CTC ACC GAA GAA AAC ACG AAA GGA CCG GGC					
---	---	---	---	---	---
Lys Asn Lys Gly Leu Tyr Leu Gly Leu Thr Glu Glu Asn Thr Lys Gly Pro Gly					
927	936	945	954	963	972
GGT GTG ACA ACA GGC CTT TCG CAC CTT GTG GAA CTC GGT GTT ACA CAC GTT CAT					
---	---	---	---	---	---
Gly Val Thr Thr Gly Leu Ser His Leu Val Glu Leu Gly Val Thr His Val His					
981	990	999	1008	1017	1026
ATA CTT CCT TTC TTT GAT TTC TAC ACA GGC GAC GAA CTC GAT AAA GAT TTC GAG					
---	---	---	---	---	---
Ile Leu Pro Phe Phe Asp Phe Tyr Thr Gly Asp Glu Leu Asp Lys Asp Phe Glu					
1035	1044	1053	1062	1071	1080
AAC TAC TAC AAC TGG GGT TAC GAT CCT TAC CTG TTC ATG GTT CCG GAG GGC AGA					
---	---	---	---	---	---
Lys Tyr Tyr Asn Trp Gly Tyr Asp Pro Tyr Leu Phe Met Val Pro Glu Gly Arg					

Figure 14 (Continued)

Thermotoga maritima Pullulanase (6GP3) (continued)

1089	1098	1107	1116	1125	1134
TAC	TCA	ACC	GAT	CCC	AAA
AAA	AAC	CCA	CAC	ACG	AGA
---	---	---	---	---	---
Tyr	Ser	Thr	Asp	Pro	Lys
1143	1152	1161	1170	1179	1188
GTC	AAA	GCC	CTT	CAC	AAA
---	---	---	---	---	---
Val	Lys	Ala	Leu	His	Lys
1197	1206	1215	1224	1233	1242
CAC	ACC	TAC	GGT	ATA	GGC
---	---	---	---	---	---
His	Thr	Tyr	Gly	Ile	Gly
1251	1260	1269	1278	1287	1296
TTC	TAC	AGA	ATC	GAC	AAG
---	---	---	---	---	---
Phe	Tyr	Arg	Ile	Asp	Lys
1305	1314	1323	1332	1341	1350
GTC	ATC	GCA	AGC	GAA	AGA
---	---	---	---	---	---
Val	Ile	Ala	Ser	Glu	Arg
1359	1368	1377	1386	1395	1404
TAC	TGG	GTA	AAG	GAG	TAT
---	---	---	---	---	---
Tyr	Trp	Val	Lys	Glu	Tyr
1413	1422	1431	1440	1449	1458
ATC	GAC	AAA	AAG	ACA	ATG
---	---	---	---	---	---
Ile	Asp	Lys	Thr	Met	Leu
1467	1476	1485	1494	1503	1512
ACT	ATC	ATT	CTC	TAC	GGC
---	---	---	---	---	---
Thr	Ile	Ile	Leu	Tyr	Gly
1521	1530	1539	1548	1557	1566
GGA	AAG	AGC	GAT	GTC	GCC
---	---	---	---	---	---
Gly	Lys	Ser	Asp	Val	Ala
1575	1584	1593	1602	1611	1620
GAC	GCA	ATA	AGG	GGT	TCC
---	---	---	---	---	---
Asp	Ala	Ile	Arg	Gly	Ser

1089 1098 1107 1116 1125 1134
TAC TCA ACC GAT CCC AAA AAC CCA CAC ACG AGA ATC AGA GAA GTC AAA GAA ATG
--- --- --- --- --- ---
Tyr Ser Thr Asp Pro Lys Asn Pro His Thr Arg Ile Arg Glu Val Lys Glu Met
1143 1152 1161 1170 1179 1188
GTC AAA GCC CTT CAC AAA CAC GGT ATA GGT GTG ATT ATG GAC ATG GTG TTC CCT
--- --- --- --- --- ---
Val Lys Ala Leu His Lys His Gly Ile Gly Val Ile Met Asp Met Val Phe Pro
1197 1206 1215 1224 1233 1242
CAC ACC TAC GGT ATA GGC GAA CTC TCT GCG TTC GAT CAG ACG GTG CCG TAC TAC
--- --- --- --- --- ---
His Thr Tyr Gly Ile Gly Glu Leu Ser Ala Phe Asp Gln Thr Val Pro Tyr Tyr
1251 1260 1269 1278 1287 1296
TTC TAC AGA ATC GAC AAG ACA GGT GCC TAT TTG AAC GAA AGC GGA TGT GGT AAC
--- --- --- --- --- ---
Phe Tyr Arg Ile Asp Lys Thr Gly Ala Tyr Leu Asn Glu Ser Gly Cys Gly Asn
1305 1314 1323 1332 1341 1350
GTC ATC GCA AGC GAA AGA CCC ATG ATG AGA AAA TTC ATA GTC GAT ACC GTC ACC
--- --- --- --- --- ---
Val Ile Ala Ser Glu Arg Pro Met Met Arg Lys Phe Ile Val Asp Thr Val Thr
1359 1368 1377 1386 1395 1404
TAC TGG GTA AAG GAG TAT CAC ATA GAC GGA TTC AGG TTC GAT CAG ATG GGT CTC
--- --- --- --- --- ---
Tyr Trp Val Lys Glu Tyr His Ile Asp Gly Phe Arg Phe Asp Gln Met Gly Leu
1413 1422 1431 1440 1449 1458
ATC GAC AAA AAG ACA ATG CTC GAA GTC GAA AGA GCT CTT CAT AAA ATC GAT CCA
--- --- --- --- --- ---
Ile Asp Lys Lys Thr Met Leu Glu Val Glu Arg Ala Leu His Lys Ile Asp Pro
1467 1476 1485 1494 1503 1512
ACT ATC ATT CTC TAC GGC GAA CCG TGG GGT GGA TGG GGA GCA CCG ATC AGG TTT
--- --- --- --- --- ---
Thr Ile Ile Leu Tyr Gly Glu Pro Trp Gly Gly Trp Gly Ala Pro Ile Arg Phe
1521 1530 1539 1548 1557 1566
GGA AAG AGC GAT GTC GCC GGC ACA CAC CTG GCA GCT TTC AAC GAT GAG TTC AGA
--- --- --- --- --- ---
Gly Lys Ser Asp Val Ala Gly Thr His Val Ala Ala Phe Asn Asp Glu Phe Arg
1575 1584 1593 1602 1611 1620
GAC GCA ATA AGG GGT TCC GTG TTC AAC CCG AGC GTC AAG GGA TTC GTC ATG GGA
--- --- --- --- --- ---
Asp Alu Ile Arg Gly Ser Val Phe Asn Pro Ser Val Lys Gly Phe Val Met Gly

Figure 14 (Continued)

Thermotoga maritima Pullulanase (6GP3) (continued)

1629	1638	1647	1656	1665	1674
GGA TAC GGA AAG GAA ACC AAG ATC AAA AGG GGT GTT GTT CGA AGC ATA AAC TAC	---	---	---	---	---
Gly Tyr Gly Lys Glu Thr Lys Ile Lys Arg Gly Val Val Gly Ser Ile Asn Tyr	---	---	---	---	---
1683	1692	1701	1710	1719	1728
GAC GGA AAA CTC ATC AAA AGT TTC GCC CTT GAT CCA GAA GAA ACT ATA AAC TAC	---	---	---	---	---
Asp Gly Lys Leu Ile Lys Ser Phe Ala Leu Asp Pro Glu Glu Thr Ile Asn Tyr	---	---	---	---	---
1737	1746	1755	1764	1773	1782
GCA GCG TGT CAC GAC AAC CAC ACA CTG TGG GAC AAG AAC TAC CTT GCC GCC AAA	---	---	---	---	---
Ala Ala Cys His Asp Asn His Thr Leu Trp Asp Lys Asn Tyr Leu Ala Ala Lys	---	---	---	---	---
1791	1800	1809	1818	1827	1836
GCT GAT AAG AAA AAG GAA TCG ACC GAA GAA CTG AAA AAC GCC CAG AAA CTG	---	---	---	---	---
Ala Asp Lys Lys Glu Trp Thr Glu Glu Leu Lys Asn Ala Gln Lys Leu	---	---	---	---	---
1845	1854	1863	1872	1881	1890
GCT GGT GCG ATA CTT CTC ACT TCT CAA CGT GTT CCT TTC CTC CAC GGA GGG CAG	---	---	---	---	---
Ala Gly Ala Ile Leu Leu Thr Ser Gln Gly Val Pro Phe Leu His Gly Gln	---	---	---	---	---
1899	1908	1917	1926	1935	1944
GAC TTC TGC AGG ACG ACG AAT TTC AAC GAC AAC TCC TAC AAC GCC CCT ATC TCG	---	---	---	---	---
Asp Phe Cys Arg Thr Thr Asn Phe Asn Asp Asn Ser Tyr Asn Ala Pro Ile Ser	---	---	---	---	---
1953	1962	1971	1980	1989	1998
ATA AAC GGC TTC GAT TAC GAA AGA AAA CTT CAG TTC ATA GAC GTG TTC AAT TAC	---	---	---	---	---
Ile Asn Gly Phe Asp Tyr Glu Arg Lys Leu Gln Phe Ile Asp Val Phe Asn Tyr	---	---	---	---	---
2007	2016	2025	2034	2043	2052
CAC AAG GGT CTC ATA AAA CTC AGA AAA GAA CAC CCT GCT TTC AGG CTG AAA AAC	---	---	---	---	---
His Lys Gly Leu Ile Lys Leu Arg Lys Glu His Pro Ala Phe Arg Leu Lys Asn	---	---	---	---	---
2061	2070	2079	2088	2097	2106
GCT GAA GAG ATC AAA AAA CAC CTG GAA TTT CTC CCG GCC GGG AGA AGA ATA GTT	---	---	---	---	---
Ala Glu Glu Ile Lys Lys His Leu Glu Phe Leu Pro Gly Gly Arg Arg Ile Val	---	---	---	---	---
2115	2124	2133	2142	2151	2160
GCG TTC ATG CTT AAA GAC CAC GCA GGT GGT GAT CCC TGG AAA GAC ATC GTG GTG	---	---	---	---	---
Ala Phe Met Leu Lys Asp His Ala Gly Gly Asp Pro Trp Lys Asp Ile Val Val	---	---	---	---	---

Figure 14 (Continued)

Thermotoga maritima Pullulanase (6GP3) (continued)

2169	2178	2187	2196	2205	2214												
ATT	TAC	AAT	CGA	AAC	TTA	GAG	AAG	ACA	ACA	TAC	AAA	CTG	CCA	GAA	GGA	AAA	TGG
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Ile	Tyr	Asn	Gly	Asn	Leu	Glu	Lys	Thr	Thr	Tyr	Lys	Leu	Pro	Glu	Gly	Lys	Trp
2223	2232	2241	2250	2259	2268												
AAT	GTG	GTT	GTG	AAC	AGC	CAG	AAA	GCC	GGA	ACA	GAA	GTG	ATA	GAA	ACC	GTC	GAA
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Asn	Val	Val	Val	Asn	Ser	Gln	Lys	Ala	Gly	Thr	Glu	Val	Ile	Glu	Thr	Val	Glu
2277	2286	2295	2304	2313	3												
GGA	ACA	ATA	GAA	CTC	GAT	CCG	CTT	TCC	GCG	TAC	GTT	CTG	TAC	AGA	GAG	TGA	***
---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---
Gly	Thr	Ile	Glu	Leu	Asp	Pro	Leu	Ser	Ala	Tyr	Val	Leu	Tyr	Arg	Glu	***	

Figure 14 (Continued)

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/00092

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :Please See Extra Sheet.

US CL :435/201, 252.33; 536/23.2

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/201, 252.33; 536/23.2

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

aps, caplus, biosis

search terms: glycosidase(s), thermococcus, staphylothermus, pyrococcus

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	VOORHORST et al. Characterization of the celB gene coding for β -glucosidase from the hyperthermophilic archaeon Pyrococcus furiosus and its expression and site-directed mutation in Escherichia coli. J. Bacteriology. December 1995, Vol. 177, No. 24, pages 7105-7111, especially pages 7105, 7106 and 7108.	1-9
Y	Database CAPLUS on STN, CAS, (Columbus, OH, USA), AN 1996:106914, KENGEN et al. "An extremely thermostable β -glucosidase from the hyperthermophilic archaeon Pyrococcus furiosus; a comparison with other glycosidases." Biocatalysis 1994, Vol. 11, No. 2, pages 79-88. Abstract.	1-9

 Further documents are listed in the continuation of Box C. See patent family annex.

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•A• document defining the general state of the art which is not considered to be of particular relevance	•T• later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
•E• earlier document published on or after the international filing date	•X• document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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•P• document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search
29 MARCH 1997

Date of mailing of the international search report

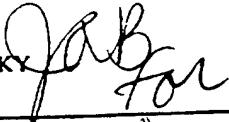
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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/00092

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X,P	BAUER et al. Comparison of β -glucosidase and β -mannosidase from the hyperthermophilic archaeon <i>Pyrococcus furiosus</i> . <i>J. Biol. Chem.</i> 27 September 1996, Vol. 271, No. 39, pages 23749-23755, see entire document.	1-9

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/00092

A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

C12N 9/26, 1/20; C07H 21/04

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

Group I, claims 1-9, drawn to a DNA, a vector comprising the DNA, a cell transformed with the same and a process for producing a peptide.

Group II, claim 10, drawn to an enzyme.

Group III, claim 11, drawn to a method of use of an enzyme.

The inventions listed as Groups I and II do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: A DNA of Group I and an enzyme of Group II are different compounds with different chemical structures and different utilities and therefore do not share a special technical feature. The method of Group III uses an enzyme and therefore does not share a special technical feature with Group I. PCT Rule 1.475(d) does not provide for the multiple products or methods within a single application and therefore unity of invention is lacking with regard to groups I, II and III.

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